

# RNA bioinformatics: Still combinatorial in 2023?

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UMR 7161 CNRS & École Polytechnique

# Who am I?

- ▶ Initial background in Computer Science
- ▶ Dabbled in Theoretical Comp Sci/Discrete Maths (random gen, disc algos)
- ▶ Contributing to RNA structural/omics bioinfo
- ▶ Cultural shock getting into Bioinformatics  
Old enough to remember the first "AI Bioinfo winter" (SVMs)

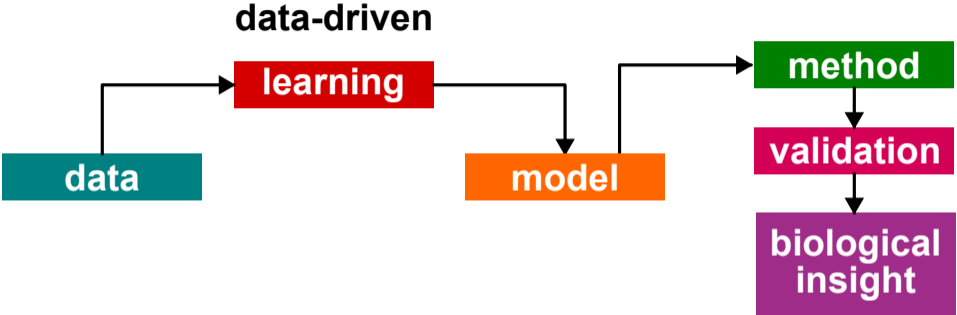
Strong interest in defining/enforcing scientific standards

- ▶ Associate editor@OUP Bioinformatics
- ▶ Proceedings chair for ISMB/ECCB 2023 (with Sushmita Roy)
- ▶ President for committee rewarding best French PhD in Computer Science

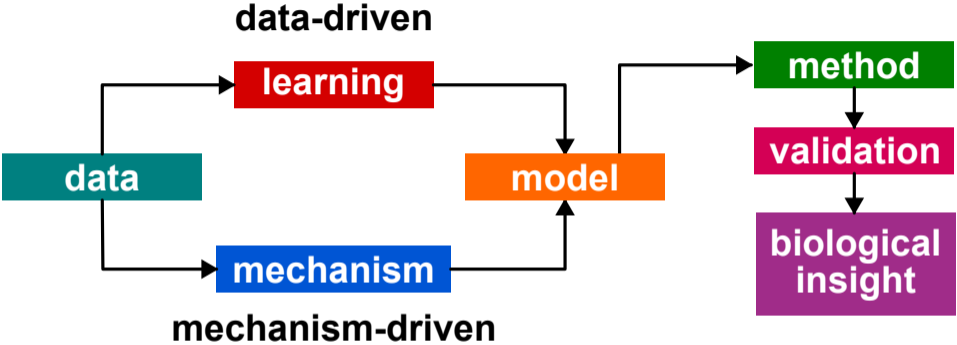
# A personal take on predictive Bioinformatics



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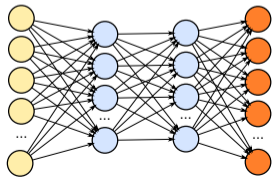
Method dev. as a modeling discipline:

Mechanism-driven model + Exact/deterministic algorithms  
→ Performance as (in)validation of model

# Machine Learning (ML): The beauty...

Machine Learning as a tool for scientific discovery

- ▶ Great promises
- ▶ Self-improving methods
- ▶ Generates/prioritizes hypotheses
- ▶ Available workforce (ubiquitous in curriculums)
- ▶ Highly promoted/funded by research institutions and glamorous journals. . .



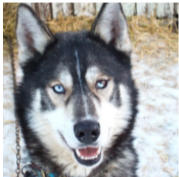
**Shut up and  
take my money**



# Machine Learning (ML): The beauty... and the beast

Multiple (potential) pitfalls for ML in Bio\*:

- ▶ Tricky evaluation (data leakage) → Extrapolation/generalization???
- ▶ Reproducibility issues (code/datasets availability, stability, retraining)
- ▶ Fishing expeditions/storytelling, selective reporting
- ▶ Educational deadend?
- ▶ Future(?) ecological disaster? Random blue checkmarks AI zealots on Twitter (grumble. . .)

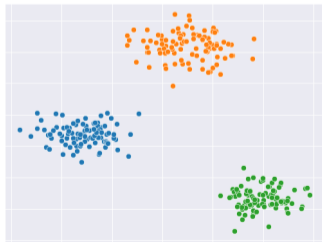


(a) Husky classified as wolf



(b) Explanation

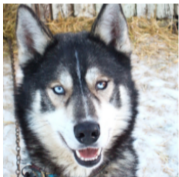
[Ribeiro et al, KDD'16]



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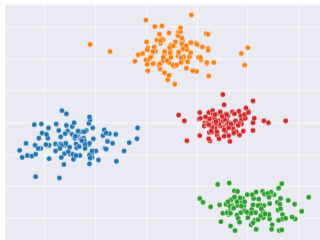


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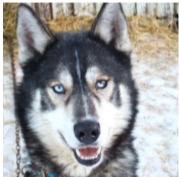




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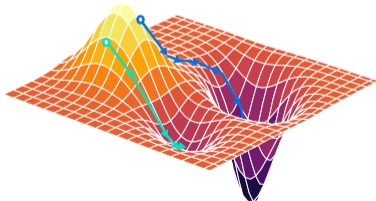
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**Available upon request**

*aka iff I'm in a good mood,  
PhD/postdoc still in lab, HDDs haven't burned,  
pharma hasn't expressed interest in data...*



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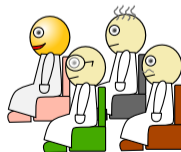
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Fifth law of thermodynamics (continued)

```
...  
-0.31622776601683794 0.31622776601683794  
-0.3157663248679193 0.3160839282916222  
0.006806069733149146 0.17777128902976705  
0.4472135954999579 1.433348584081719  
-1.5736761136523203 1.433348584081719  
-0.0002340648727882 0.4522609460629265  
...
```

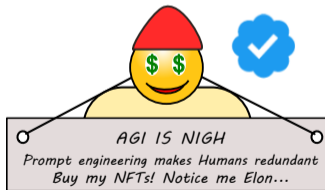
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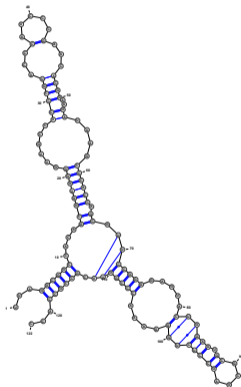


# RNA structure(s)

RNA = Linear Polymer = Nucleotides sequence  $w \in \{A, C, G, U\}^*$

```
UUAGGCGGCCACAGC
GGUGGGGUUGCCUCC
CGUACCCAUCCCGAA
CACGGAAGAUAAAGCC
CACCAGCGUUCGGG
GAGUACUGGAGUGCG
CGAGCCUCUGGGAAA
CCCGGUUCGCCGCCA
CC
```

Primary struct.



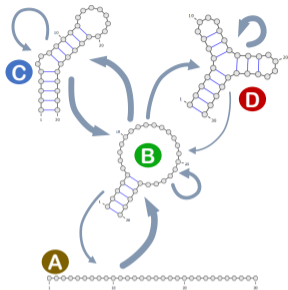
Secondary (2D) struct.



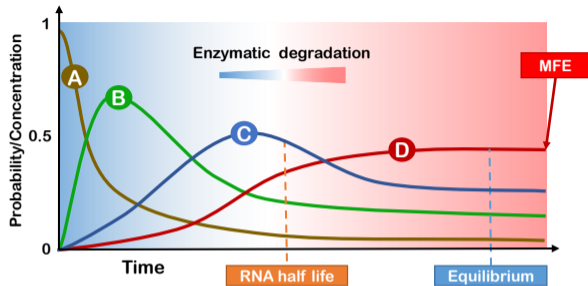
Tertiary ( $\approx$  3D) struct.

5s rRNA (PDBID: 1K73:B)

# Paradigms in RNA structural bioinformatics



A – Kinetic Landscape  
Continuous-time Markov chain



B – Evolution of concentrations

**Free-energy**  $E : \Sigma^* \times \mathcal{S} \rightarrow \mathbb{R}$ , at Boltzmann equilibrium  $\mathbb{P}(S | w) \propto e^{-E(w,S)/RT}$

- ▶ **Minimum Free-Energy (MFE)**: Functional structure = Most stable/probable
- ▶ **Partition function**: Equilibrium properties of Boltzmann ensemble
- ▶ **Kinetics**: Finite-time evolution of concentrations/probabilities

# A crowded ML field for RNA 2D prediction



Method	Output	PKs?	Architecture	Availability
CONTRAFold	Pairwise contacts	No	CLLM	Code+weights+webserver
EternaFold	Pairwise contacts	No	CLLM	Code+weights+webserver
DMfold	DBN	Yes	bi-LSTM	Code only
RNA-state-inf	Binary paired/unpaired	N/A	bi-LSTM	Code only
SPOT-RNA2	Pairwise contacts	Yes	CNN	Code+weights+webserver
CROSS	Binary paired/unpaired	N/A	CNN-like	Webserver
RPRes	Binary paired/unpaired	N/A	bi-LSTM+CNN	Code only
2dRNA	Pairwise contacts	Yes	bi-LSTM+CNN	Webserver
2dRNA-LD	Pairwise contacts	Yes	bi-LSTM+CNN	Webserver
SPOT-RNA	Pairwise contacts	Yes	CNN+bi-LSTM	Code+weights+webserver
MXfold2	Pseudo-dG	No	CNN+bi-LSTM	Code+weights+webserver
CNNFold	Pairwise contacts	Yes	CNN(NxN input)	Code+weights
UFold	Pairwise contacts	Yes	CNN(NxN input)	Code+weights+webserver
CDPFold	DBN	No	CNN(NxNinput)	Code
E2EFold	Pairwise contacts	Yes	Transformer+CNN	Code+weights
ATTFold	Pairwise contacts	Yes	Transformer+CNN	No

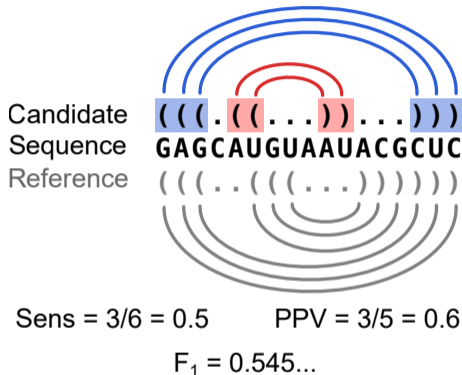
[Wu *et al*, Briefings in Bioinfo 2023]

# Performances of 2D structure prediction

## RNAstrand benchmark

[Adronescu *et al*, BMC Bioinf 2008]

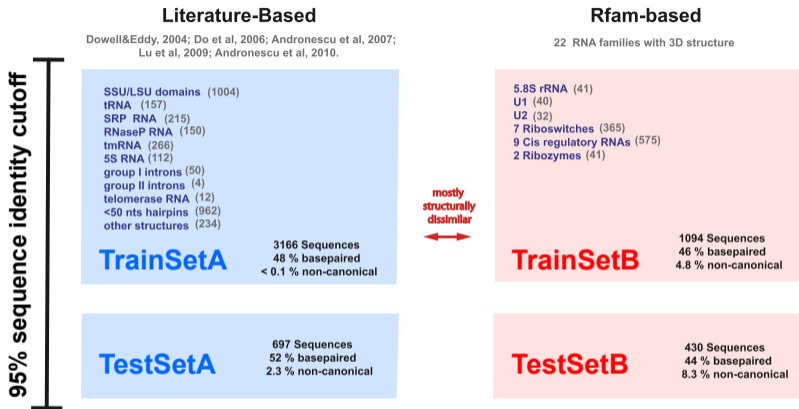
Method	F <sub>1</sub>
RNAfold 1.8.5	0.737
UNAFold 3.8	0.725
RNAstructure 5.7	0.744



$$F_1\text{-score} = \frac{2 \times \text{PPV} \times \text{Sens}}{\text{PPV} + \text{Sens}}$$



# The TORNADO dataset



[Rivas *et al*, RNA 2012]

TrainSetA vs TestSetA: 95% sim. cutoff → Learn  $k$ -mer to template association

(May happen even for extreme cutoffs)

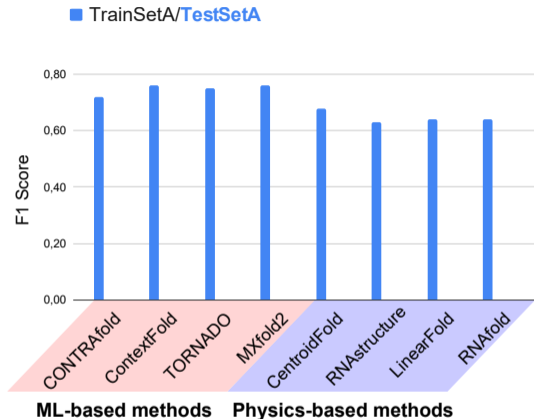
TrainSetA vs TestSetB: Rewards learning structurally generalizable models

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[Sato *et al*, Nature Comm 2021]

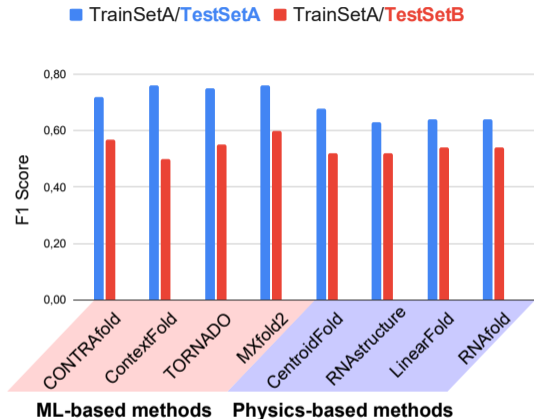
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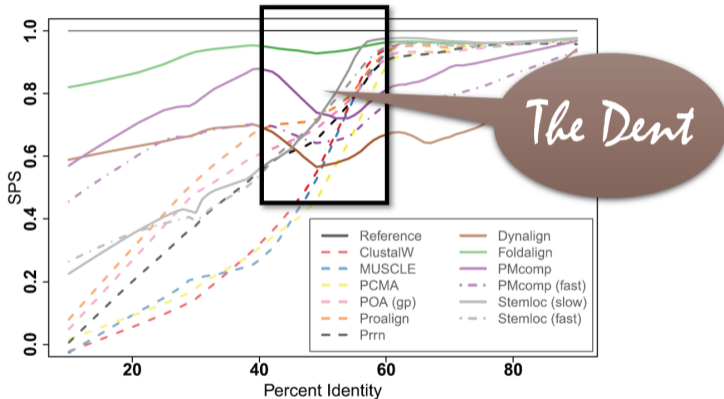


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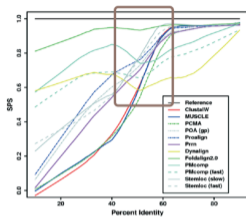
# Biased benchmarks: precedent in comparative folding/alignment

**Bralibase:** Benchmark for comp. RNA folding [Gardner,Wilm & Washietl, NAR 2005]

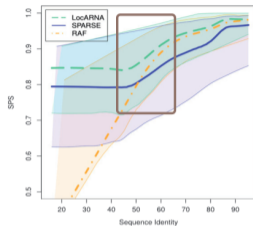


[Löwes *et al*, Briefings in Bioinfo 2016]

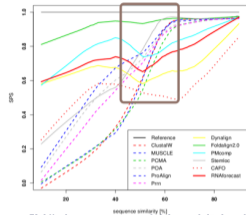
# Biased benchmarks: precedent in comparative folding/alignment



[Gardner *et al*, NAR 2005]



[Will *et al*, Bioinformatics 2015]



[Höchsmann *et al*, Unpublished]

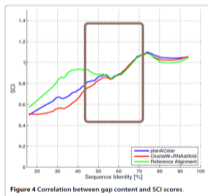
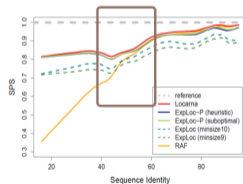
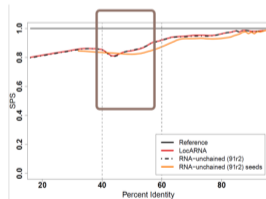


Figure 4 Correlation between gap content and SCS scores.

[Bremges *et al*, BMC Bioinfo, 2010]



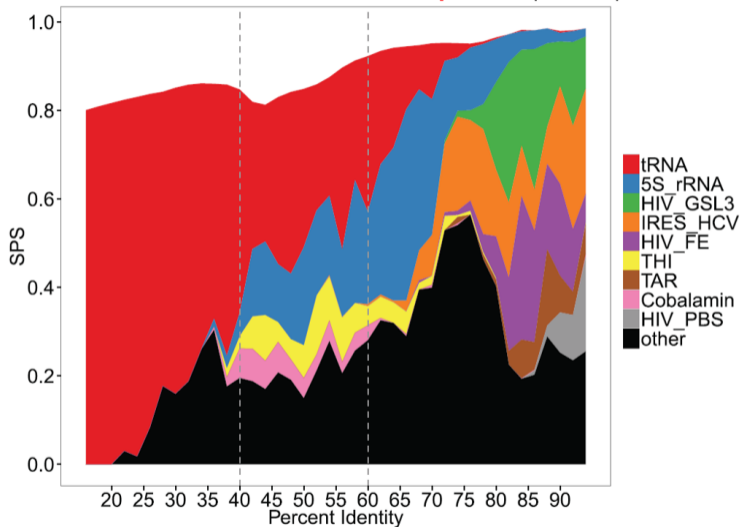
[Schmiedl *et al*, RECOMB 2012]



[Bourgeade *et al*,] Comp Biol, 2015]

[Löwes *et al*, Briefings in Bioinfo 2016]

# Biased benchmarks: precedent in comparative folding/alignment



[Löwes *et al*, Briefings in Bioinfo 2016]

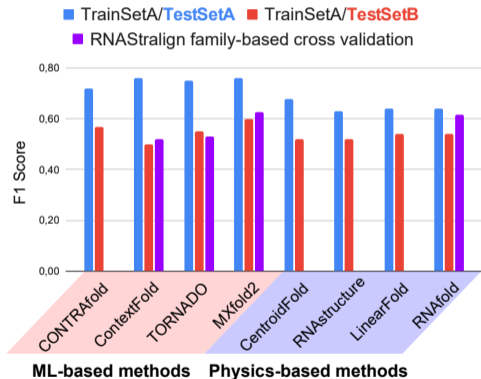
# The (nc)RNA datasphere

- ▶ 34M sequences, inc 22M presumably structured (RNACentral)
- ▶ 4000+ functional ncRNA families (RFAM)
- ▶ 250-300 non-redundant 3D models (PDB)

Existing methods trained on datasets:

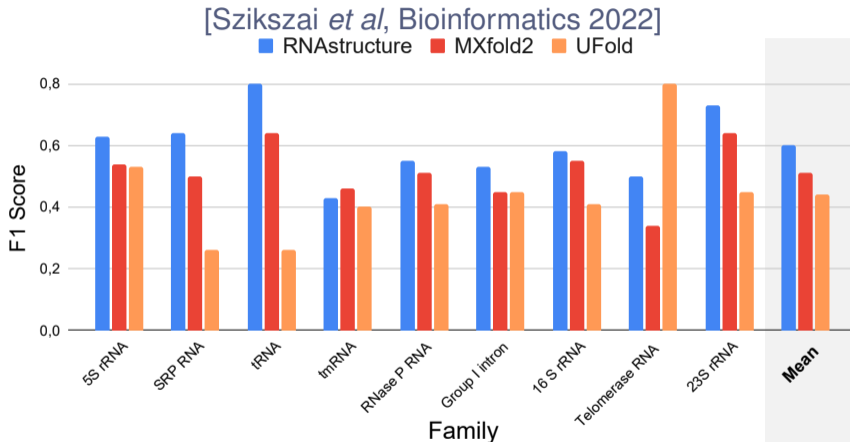
- ▶ highly-redundant sequence-wise
- ▶ low-diversity structure-wise

Do ML methods generalize to new structures?  
(Do ML perms translate into *new* biological insight?)



[Sato *et al*, Nature Comm 2021]

# Generalization to new families/structures remains problematic



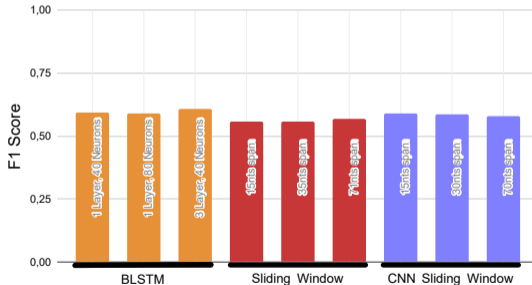
Family-fold cross-validation on **Archivell** dataset [Sloma & Mathews, RNA 2016]  
3974 RNAs of length 77-438 (large rRNAs split into smaller domains)



# What if you had access to (unbounded) additional data?

Idea: Assess NN models' capacity to emulate RNAfold on random sequences

[Flamm *et al*, Frontiers in Bioinfo 2022]

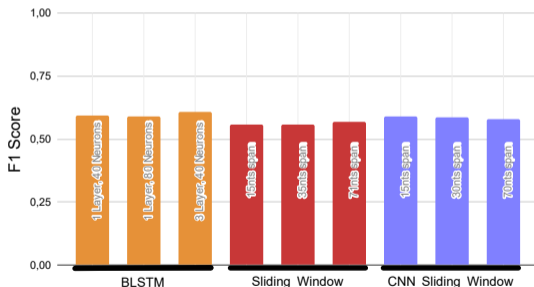


Perfs *plateau* at 80k seq/structs (70nts)

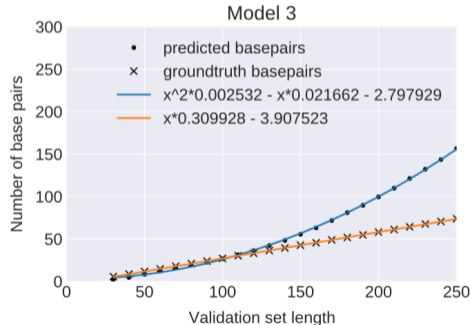
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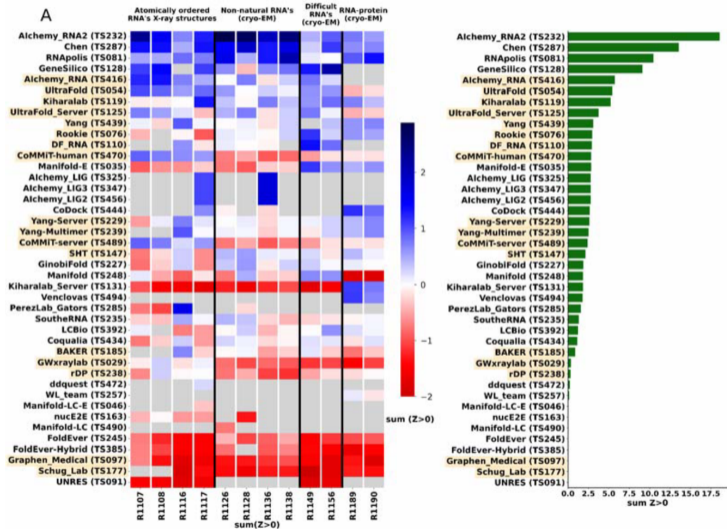


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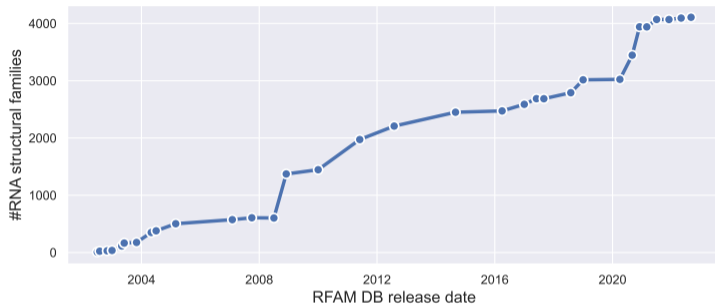
Popular CNN predicts  $\Theta(n^2)$  BPs!

# RNA 3D structure: No AlphaFold moment at CASP15



[Das *et al*, under review]

# Conclusions and musings



- ▶ Still a need for improved RNA prediction (possibly ML-based)
- ▶ Purely combinatorial methods still  $\pm$  state-of-the-art for new families. . .
- ▶ Hybrid approaches *à la* MxFold2: Best of both worlds?
- ▶ Assessing intrinsic limits of architectures: RNAFold as surrogate model

# Conclusions and musings

So what's special about RNA?

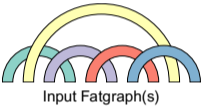
- ▶ Modular but combinatorial structure
- ▶ New folds being routinely discovered (+ can be designed)
- ▶ Relatively scarce 3D data
- ▶ Opportunity: Tons of probing data (ML)
- ▶ Potential of LLMs/transformers (incoming)
- ▶ Pseudoknots-ready algorithms

# Automated derivation of folding algorithms inc. pseudoknots

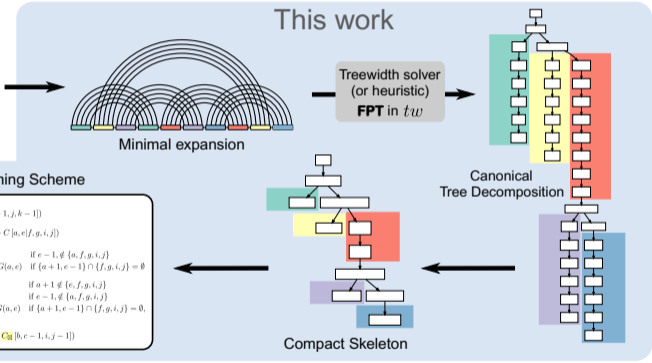
PK pattern(s) of interest (e.g. 3D models)



Abstracted as ↓



Input Fatgraph(s)



Dynamic Programming Scheme

$$\begin{aligned}
 A &= \min_{a,g,h,j,k} (B[a,g,h,j] + C[a][g,h-1,j,k-1]) \\
 B[a,g,h,j] &= \min_{e,f,i} (C[a][e,f-1,h,i-1] + C[a,e][f,g,i,j]) \\
 C[a,e][f,g,i,j] &= \min \begin{cases} C^*[a,e-1][f,g,i,j], & \text{if } e-1 \notin \{a,f,g,i,j\} \\ C[a+1,e-1][f,g,i,j] + \Delta G(a,e), & \text{if } \{a+1,e-1\} \cap \{f,g,i,j\} = \emptyset \end{cases} \\
 C[a,e][f,g,i,j] &= \min \begin{cases} C^*[a+1,e][f,g,i,j], & \text{if } a+1 \notin \{e,f,g,i,j\} \\ C^*[a,e-1][f,g,i,j], & \text{if } e-1 \notin \{a,f,g,i,j\} \\ C[a+1,e-1][f,g,i,j] + \Delta G(a,e), & \text{if } \{a+1,e-1\} \cap \{f,g,i,j\} = \emptyset, \\ D^*[a,e+1][f,g,i,j] \end{cases} \\
 D[b,d,f,g,i,j] &= \min_c (C[a][c,d-1,f,g-1] + C[a][b,c-1,i,j-1])
 \end{aligned}$$

⚙️ General algorithm for Fatgraph MFE folding problem  
 Correct for any input sequence (+ C/C++ Code generation)

Complexity:  $O(n^{tw})$  or  $O(n^{tw+1})$  for simple energy models  
 $O(n^{tw+1})$  for full Turner model

[Marchand *et al*, WABI 2023]

# Conclusions and musings

(RNA) community needs to enforce stricter standards for ML publications:

- ▶ Enforce datasets and source code availability  
[Szikszai *et al*, *Bioinfo'22*] found 4/8 recent DL methods non-functional
- ▶ Realistic retraining mandatory  
Precondition for self-improvement, benchmarking of novel methods
- ▶ Consider mechanistic and ML methods as largely incomparable
- ▶ Better datasets/benchmarks needed, but perhaps not sufficient
- ▶ Sequence-based leakage should be systematically investigated

# What are ze questions?



Many thanks to:

- ▶ Ze **SFBI** for putting ze session together
- ▶ Ze **ISCB** (Diane and Steven)
- ▶ Ze whole **proceedings** program committee (Sushmita + 20<sup>+</sup> ACs + 200<sup>+</sup> members)
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