



# Goals

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- ▶ To help you survive the *RNA data jungle*.
- ▶ To conceptually and practically connect the three levels of RNA structural information.
- ▶ To introduce mature prediction and annotation tools.
- ▶ To illustrate the structure-informed curation RNA alignments.
- ▶ To keep this fun and interactive.



# Schedule (French)

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When?	What?
9:30	Introduction
9:45	First session: Databases, 2D structure prediction tools, 3D annotations tools, hands on.
10:30	Interactive coffee break
10:45	Second session: Ensemble approaches, comparative methods, further refinement of alignments, assessment.
12:30	Discussion
13:00	Lunch

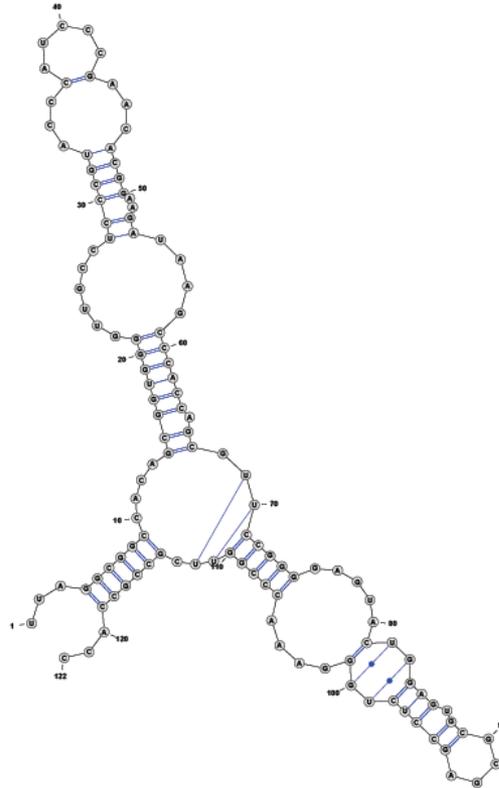


# RNA structure(s)

---

```
UUAGGCGGCCACAGC
GGUGGGGUUGCCUCC
CGUACCCAUCCCGAA
CACGGAAGAUAAAGCC
CACCAGCGUUCCGGG
GAGUACUGGAGUGCG
CGAGCCUCUGGGAAA
CCCGGUUCGCCGCCA
CC
```

Primary structure

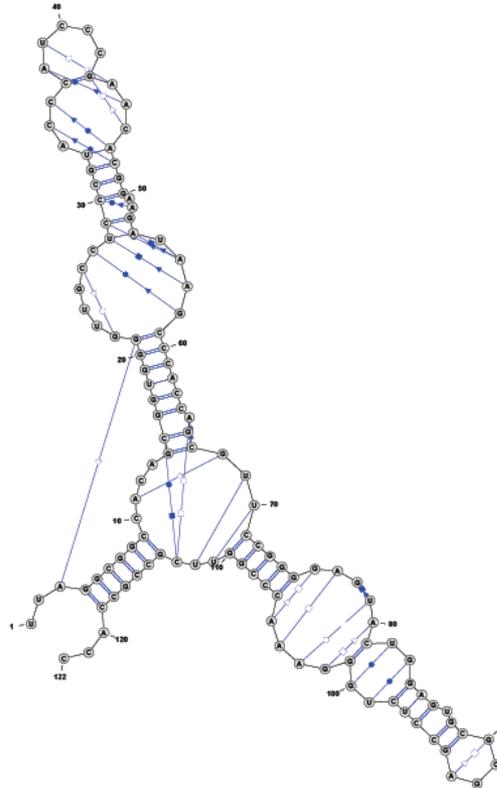


# RNA structure(s)

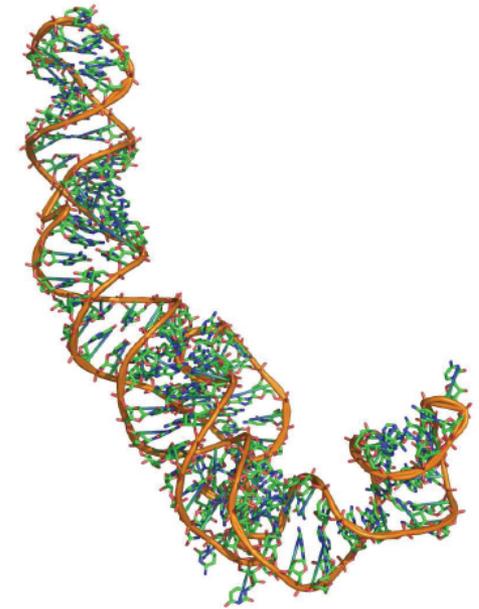
---

```
UUAGGCGGCCACAGC
GGUGGGGUUGCCUCC
CGUACCCAUCCCGAA
CACGGAAGAUAAAGCC
CACCAGCGUUCGGG
GAGUACUGGAGUGCG
CGAGCCUCUGGGAAA
CCGGUUCGCCGCCA
CC
```

Primary structure



Secondary<sup>+</sup> structure



Tertiary structure

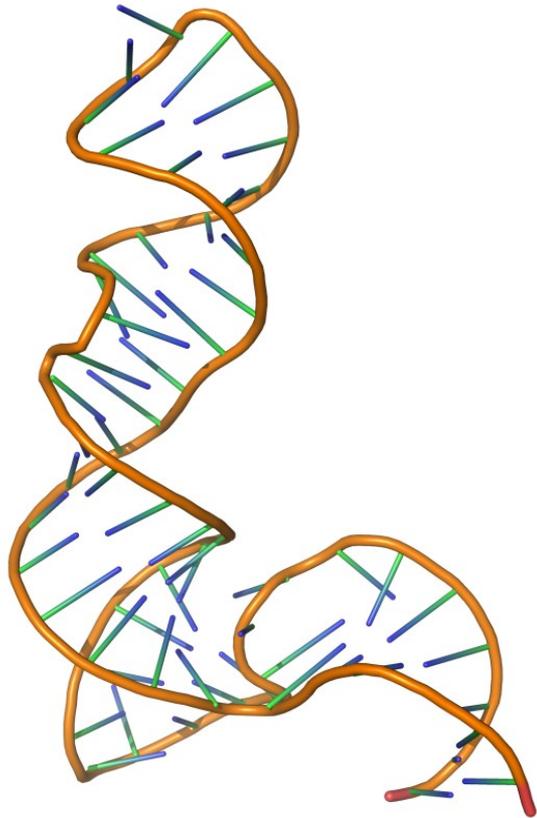
Source: 5s rRNA (PDBID: 1K73:B)

---

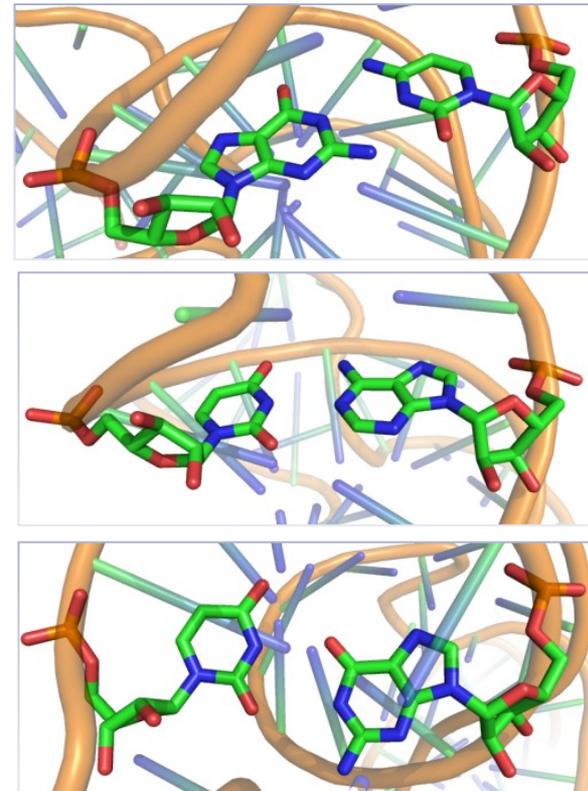


# How RNA folds

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5s rRNA (PDB ID: 1UN6)



G/C

U/A

U/G

Canonical base-pairs

RNA folding = Hierarchical stochastic process driven by/resulting in the pairing (hydrogen bonds) of a subset of its bases.

---



# Sources of RNA data

Name	Data type	Scope	Description	File formats	#Entries	URL
<b>PDB</b>	All-atoms	General	RCSB Protein Data Bank – Global repository for 3D molecular models	PDB	~1,900 models	<a href="http://www.pdb.org">http://www.pdb.org</a>
<b>NDB</b>	All-atoms, Secondary structures	General	Nucleic Acids Database – Nucleic acids models and structural annotations.	PDB, RNAML	~2,000 models	<a href="http://bit.ly/rna-ndb">http://bit.ly/rna-ndb</a>
<b>RFAM</b>	Alignments, Secondary structures <sup>3</sup>	General	RNA FAMILies – Multiple alignments of RNA as functional families. Features consensus secondary structures, either predicted and/or manually curated.	STOCKHOLM, FASTA	~1,973 Alignments/structures, 2,756,313 sequences	<a href="http://bit.ly/rfam-db">http://bit.ly/rfam-db</a>
<b>STRAND</b>	Secondary structures	General	The RNA secondary STRucture and statistical ANalysis Database – Curated aggregation of several databases	CT, BPSEQ, RNAML, FASTA, Vienna	4,666 structures	<a href="http://bit.ly/ssstrand">http://bit.ly/ssstrand</a>
<b>PseudoBase</b>	Secondary structures	Pseudoknotted RNAs	PseudoBase – Secondary structure of known pseudoknotted RNAs.	Extended Vienna RNA	359 structures	<a href="http://bit.ly/pkbase">http://bit.ly/pkbase</a>
<b>CRW</b>	Sequence alignments, Secondary structures	Ribosomal RNAs, Introns	Comparative RNA Web Site – Manually curated alignments and statistics of ribosomal RNAs.	FASTA, ALN, BPSEQ	1,109 structures, 91,877 sequences	<a href="http://bit.ly/crw-rna">http://bit.ly/crw-rna</a>



# RNA file formats: Sequences (alignments)

---

```
>O.sativa.1 AJ489954.1/1-104
.....UGGCUGUGACGACUAGGUGAAAUU.CAAGCUCAACAGACCAAUUCACAGGUCUC
..UCUCCAAGGCCUU.UGGAGAUUGGGAUCUGUAUGCCGA.....GU..UUCCGCUC....
.AGCCG.....
>O.sativa.2 AY013245.2/61987-62105
...GAUGGCAGUGACGACUUGGUAUAUU.CAAGCUCAACAGACCAAUUCACAGGUCUU
CCUCUCUGGAUCCAC..UCCUCUGGGAUUGAUUUG..UAUGCCGAUUUCCCGCUGAACC
GAGCCAUC....
>O.sativa.3 AJ307928.1/3-121
...GAUGGCAGUGACGACCUUGGUAUAUU.CAAGCUCAACAGACCAAUUCACAGGUCUU
..UCUCUCUGGAUCUACUCCUCAGGGAUUGAUUUG..UAUGCCGAUUUCCCGCUGAACC
GAGCCAUC....
```

## Fasta

### CLUSTAL 2.1 multiple sequence alignment

```
M.musculus.1      UGGCCUCGUUCAAGUAAUCCAGGAUAGG--CU--GUG-CAGGUCCCAAGGGCCUAUUUCU 55
H.sapiens.2      UGGCCUCGUUCAAGUAAUCCAGGAUAGG--CU--GUG-CAGGUCCCAU-GGCCUAU-CU 53
H.sapiens.3      GGACCCAGUUCAAGUAAUUCAGGAUAGGUUGU--GUG-CUGU--CCAG----CCUGUUCU 51
T.rubripes.1     CAACCGGUUCAAGUAAUCCAGGAUAGGCUCU--GUAUCUGU--CUUGG---CCUAUGCU 53
H.sapiens.1     UGGCUGGAUUCAAGUAAUCCAGGAUAGGCUGUUUCCAUCUGU--G-AGG---CCUAUUUCU 54
..*      .***** ***** *      . * *      .   ***.* **

M.musculus.1      UGGUUACU---UGCACGGGGAC 74
H.sapiens.2      UGGUUACU---UGCACGGGGAC 72
H.sapiens.3      CCAUUACU--UGGCUCGGGGAC 71
```

## Aln

---



# RNA file formats: Sequences (alignments)

---

```
# STOCKHOLM 1.0
#=GF ID      mir-22
#=GF AC      RF00653
...
O.laticipes.1      CGUUG.CCUCACAGUCGUUCUUA.CUGGCU.AGCUUUUAUGUCCCACG..
Gasterosteus_aculeat.1  GGCUG.ACCUACAGCAGUUCUUA.CUGGCA.AGCUUUUAUGUCCUCAUCU
R.esox.1           AGCUGAGCACA...CAGUUCUUA.CUGGCA.GCCUUAAGGUUUCUGUAG
...
#=GC SS_cons      .<<<<. <<. <<<<<<<<<<<<<<<<<<. <<<<. <<<<<<<. <<.....
#=GC RF           gGccg.acucaCagcaGuuCuuCa.cuGGCA.aGCuuuAuguccuuauaa

O.laticipes.1      CCCCACCGUAAAGCU.GC.CAGUUGAAGAGCUGUUGUG..UGUAACC
Gasterosteus_aculeat.1  ACCAGC..UAAAGCU.GC.CAGCUGAAGAACUGUUGUG..GUCGGCA
R.esox.1           ACAGGC..UAAACCU.GC.CAGCUGAAGAACUGCUCUG..GCCAGCU
...
#=GC SS_cons      .....>>.>>>>>>>>.>>.>>.>>>>>>>>>>>>>>>>>>.....
#=GC RF           acaaac..UaaaGCu.GC.CaGuuGaaGaaCugcuGug..gucggCu
//
```

Stockholm (RFAM)



# RNA file formats: Secondary Structures

---

```
> Rat Alanine tRNA
GAGGAUUUAGCUUAAUUAAAGCAGUUGAUUUUGCAUUUAAACAGAUGUAAGAUUAGUCUUACAGUCCUUA
(((((((...(((.....))))).((((.....)))))...(((((((...)))))))))))).
```

## Vienna (RNAFold)

```
      1590      1600      1610      1620      1630
#      |123456789|123456789|123456789|123456789|123456
$ 1590 AAAAAACUAAUAGAGGGGGACUUAGCGCCCCCAAACCGUAACCCC=1636
% 1590 ::::::::::::::: [[[[[[[:::(([]]]]]]::~))]::~:::
```

## PseudoBase



# RNA file formats: Secondary Structures

---

## BPSeq (CRW)

```
Filename: AM286415_b.bpseq
Organism: Yersinia enterocolitica subsp. enterocolitica 8081
Accession Numbers: AM286415
Citation and related information available at http://www.rna.ccbb.utexas.edu
1 U 0
...
117 U 0
118 U 236
119 G 235
120 C 234
121 C 233
122 U 232
123 G 231
124 G 230
...
230 C 124
231 C 123
232 A 122
233 G 121
234 G 120
235 C 119
236 A 118
...
```

dG = -33.48 [Initially -35.60]					
80					
1	U	0	2	80	1
2	G	1	3	79	2
3	G	2	4	78	3
4	G	3	5	77	4
5	A	4	6	76	5
6	U	5	7	0	6
7	G	6	8	75	7
...					
75	U	74	76	7	75
76	U	75	77	5	76
77	C	76	78	4	77
78	C	77	79	3	78
79	U	78	80	2	79
80	A	79	0	1	80

## CT (MFold)



# RNA file formats: Secondary Structures

---

```
HEADER      RNA                               27-JUL-09   3IGI
TITLE      TERTIARY ARCHITECTURE OF THE OCEANOBACILLUS IHEYENSIS GROUP
TITLE      2 II INTRON
COMPND     MOL_ID: 1;
COMPND     2 MOLECULE: GROUP IIC INTRON;
COMPND     3 CHAIN: A;

...

ATOM       8009  P      U A 375      19.076  79.179 370.688  1.00 66.25      P
ATOM       8010  OP1    U A 375      18.815  77.862 371.313  1.00 83.22      O
ATOM       8011  OP2    U A 375      19.869  80.203 371.409  1.00 56.32      O
...
CONNECT   8654 8520
CONNECT   8655 8521
CONNECT   8658 8531
MASTER           717      0 66      0 0 0 69      6 8656      2 123 33
END
```

**PDB**



# RNA file formats: Secondary Structures

```
<?xml version="1.0"?>
<!DOCTYPE rnaml SYSTEM "rnaml.dtd">
<rnaml version="1.0">
  <molecule id="xxx">
    <sequence> ... </sequence>
    <structure> ... </structure>
  </molecule>
  <interactions> ... </interactions>
</rnaml>
```

RNAML

# RNA file formats: Secondary Structures

```
<?xml version="1.0"?>
<!DOCTYPE rnaml SYSTEM "rnaml.dtd">
<rnaml version="1.0">
  <molecule id="xxx">
    <sequence>
      <numbering-system id="1" used-in-file="false">
        <numbering-range>
          <start>1</start><end>387</end>
        </numbering-range>
      </numbering-system>
      <numbering-table length="387">
        2 3 4 5 6 7 8...
      </numbering-table>
      <seq-data>
        UGUGCCCGGC AUGGGUGCAG UCUAUAGGGU...
      </seq-data>
      ...
    </sequence>
    <structure> ... </structure>
  </molecule>
  <interactions> ... </interactions>
</rnaml>
```

RNAML

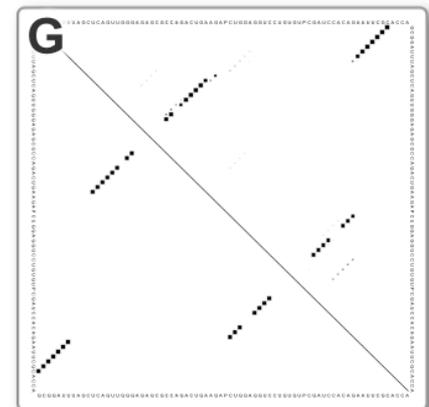
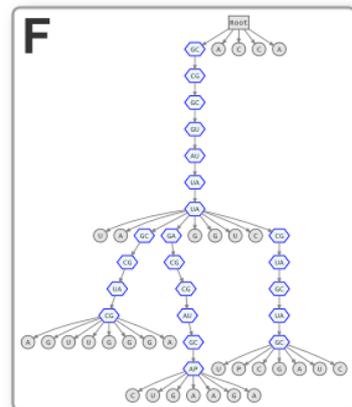
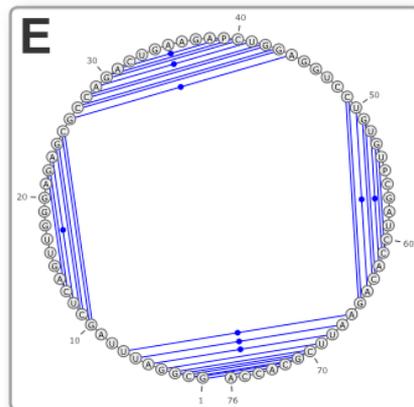
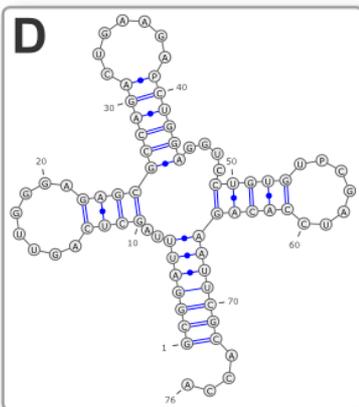
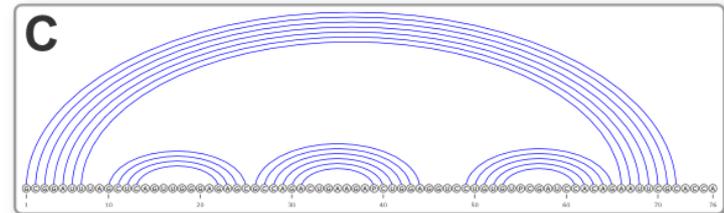
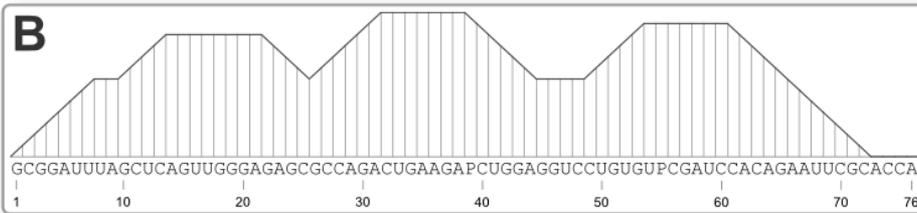
# RNA file formats: Secondary Structures

```
<?xml version="1.0"?>
<!DOCTYPE rnaml SYSTEM "rnaml.dtd">
<rnaml version="1.0">
  <molecule id="xxx">
    <sequence> ... </sequence>
    <structure>
      <model id="yyy">
        <base> ... </base> ...
        <str-annotation>
          ...
          <base-pair>
            <base-id-5p><base-id><position>2</position></base-id></base-id-5p>
            <base-id-3p><base-id><position>260</position></base-id></base-id-3p>
            <edge-5p>+</edge-5p>
            <edge-3p>+</edge-3p>
            <bond-orientation>c</bond-orientation>
          </base-pair>
          <base-pair comment="?">
            <base-id-5p><base-id><position>4</position></base-id></base-id-5p>
            <base-id-3p><base-id><position>259</position></base-id></base-id-3p>
            <edge-5p>S</edge-5p>
            <edge-3p>W</edge-3p>
            <bond-orientation>c</bond-orientation>
          </base-pair>
          ...
        </str-annotation>
      </model>
    </structure>
  </molecule>
  <interactions> ... </interactions>
</rnaml>
```

RNAML

# Secondary Structure representations

**A** (((((((((..(((.....))))((((.....)))))).....((((.....))))))))).....  
GCGGAUUUAGCUCAGUUGGGAGAGCGCCAGACUGAAGAPCUGGAGGUCCUGUGUPCGAUCCACAGAAUUCGCACCA



<http://varna.lri.fr>

# First contact

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- ▶ Run the web start version of VARNA at:

<http://varna.lri.fr/downloads.html>

- ▶ Locate and save on disk a bunch of secondary structures from the RNA Strand Database (CT or BPseq):

<http://www.rnasoft.ca/strand/>

- ▶ Load these files and using the **region highlight** feature of VARNA, highlight a region of interest.

Menu ▶ Edit ▶ Annotation ▶ New ▶ Region



# Basic prediction

Minimal free-energy folding

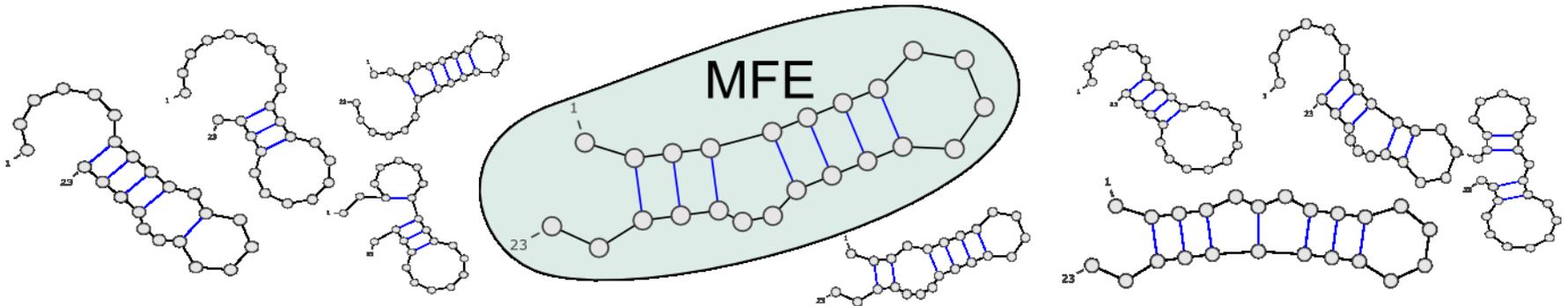
# Minimal Free-Energy (MFE) Folding

---

...CAGUAGCCGAUCGCAGCUAGCGUA...



RNAFold

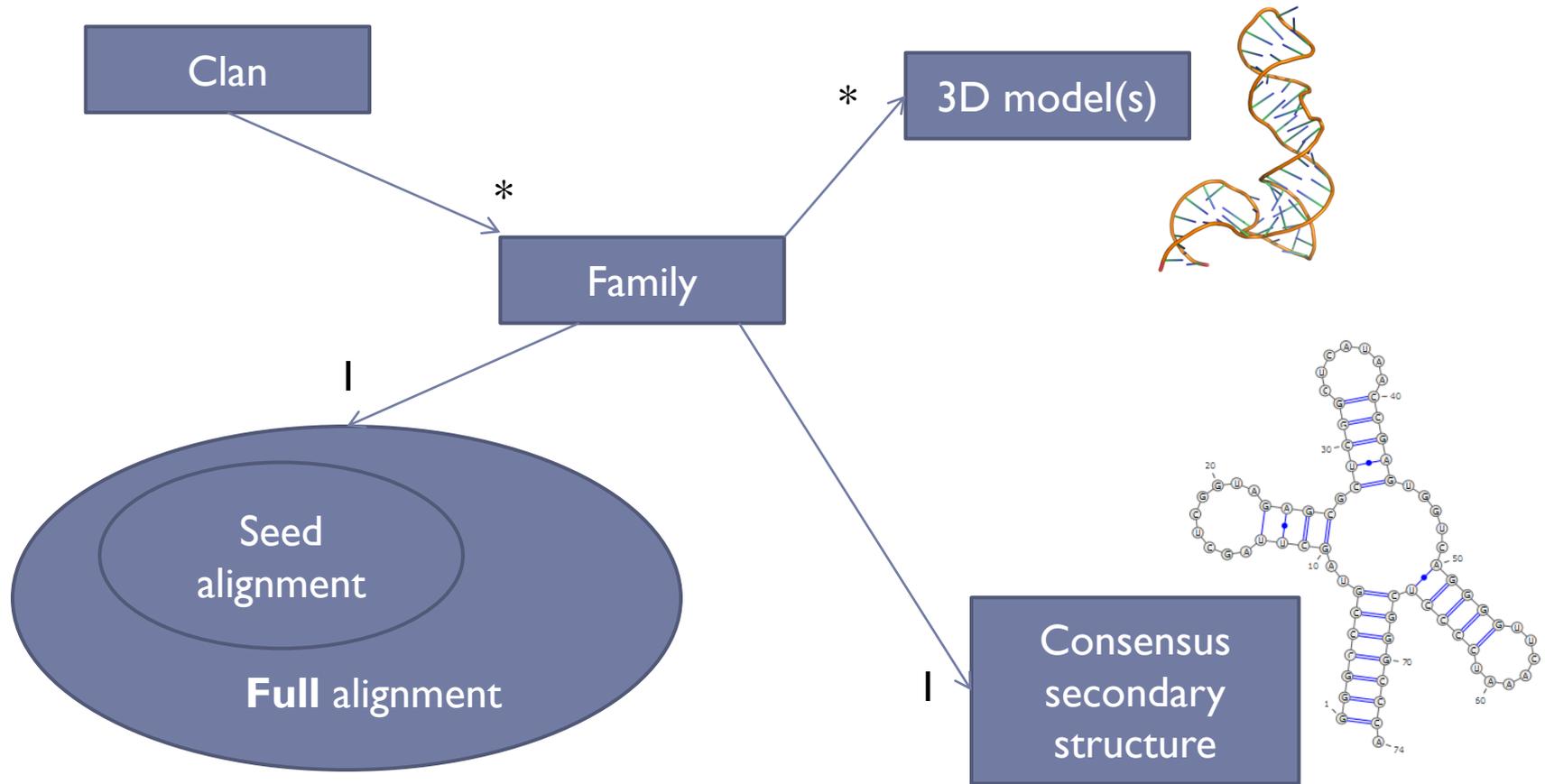


- ▶ Turner model associates energy to each compatible secondary structure.
- ▶ Vienna RNA package implements a  $O(n^3)$  algorithm for computing the most stable folding...
- ▶ ... but also offers nice visualization features.



# RFAM: RNA functional families

<http://rfam.sanger.ac.uk/>



# Minimal Free-Energy folding of RNA

---

- ▶ Get the RFAM alignment for the the DI-D4 domain of the Group II intron (RFAM ID: RF02001 – Seed – Stockholm format)  
<http://rfam.sanger.ac.uk/>
- ▶ Load the *A. Capsulatum* (*Acidobacterium\_capsu.1*) sequence in VARNA.
- ▶ Run RNAFold on this sequence using the Vienna RNA web tools suite:  
<http://rna.tbi.univie.ac.at/>
- ▶ Retrieve the result (Vienna format) and compare it with the consensus structure.
- ▶ Rerun RNAFold using more recent energy parameters  
(*Show advanced options* → *Turner 2004 energy model*)
- ▶ Compare the predictions in both models.

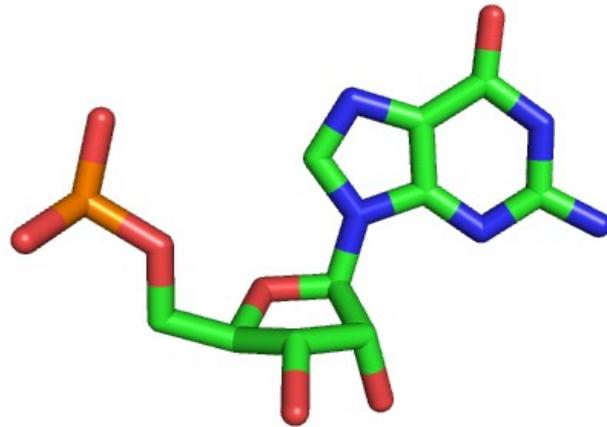


# Advanced structural features

Tertiary motifs and pseudoknots

# Non canonical interactions

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RNA nucleotides bind through edge/edge interactions.

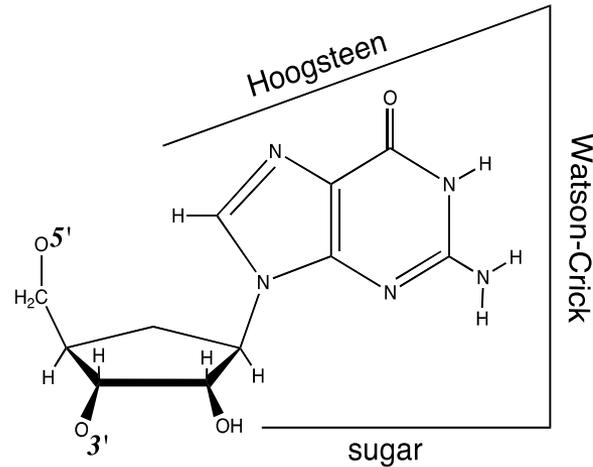
Non canonical are weaker, but cluster into modules that are **structurally constrained**, **evolutionarily conserved**, and functionally essential.

---



# Non canonical interactions

---



RNA nucleotides bind through edge/edge interactions.

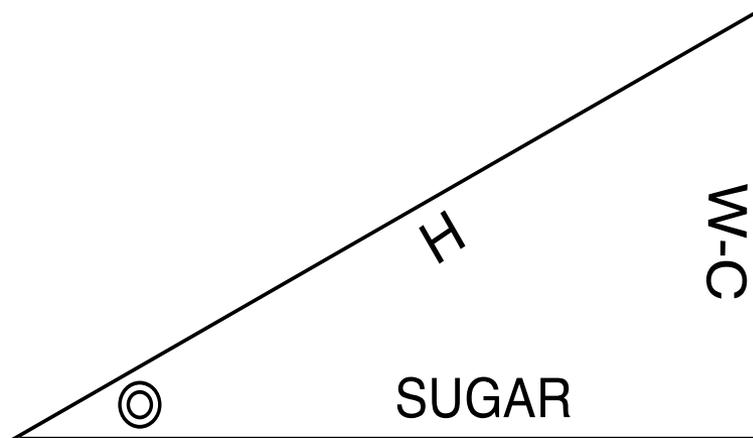
Non canonical are weaker, but cluster into modules that are **structurally constrained**, **evolutionarily conserved**, and functionally essential.

---



# Non canonical interactions

---



RNA nucleotides bind through edge/edge interactions.

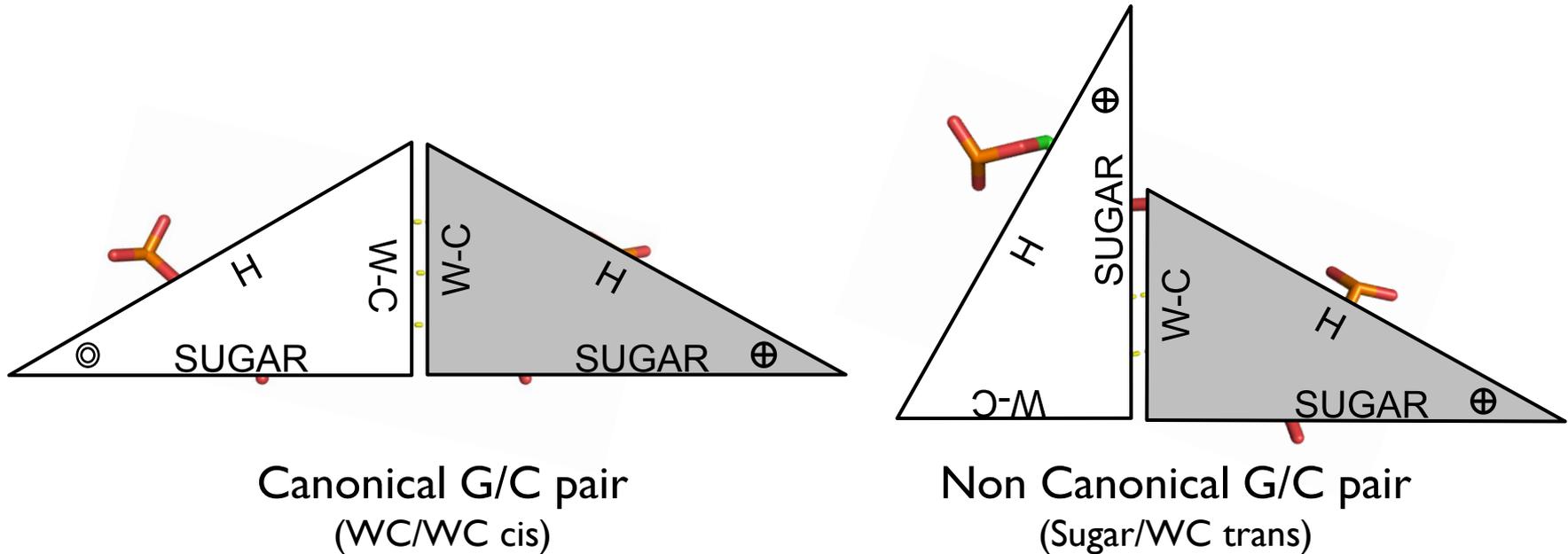
Non canonical are weaker, but cluster into modules that are **structurally constrained**, **evolutionarily conserved**, and functionally essential.

---



# Non canonical interactions

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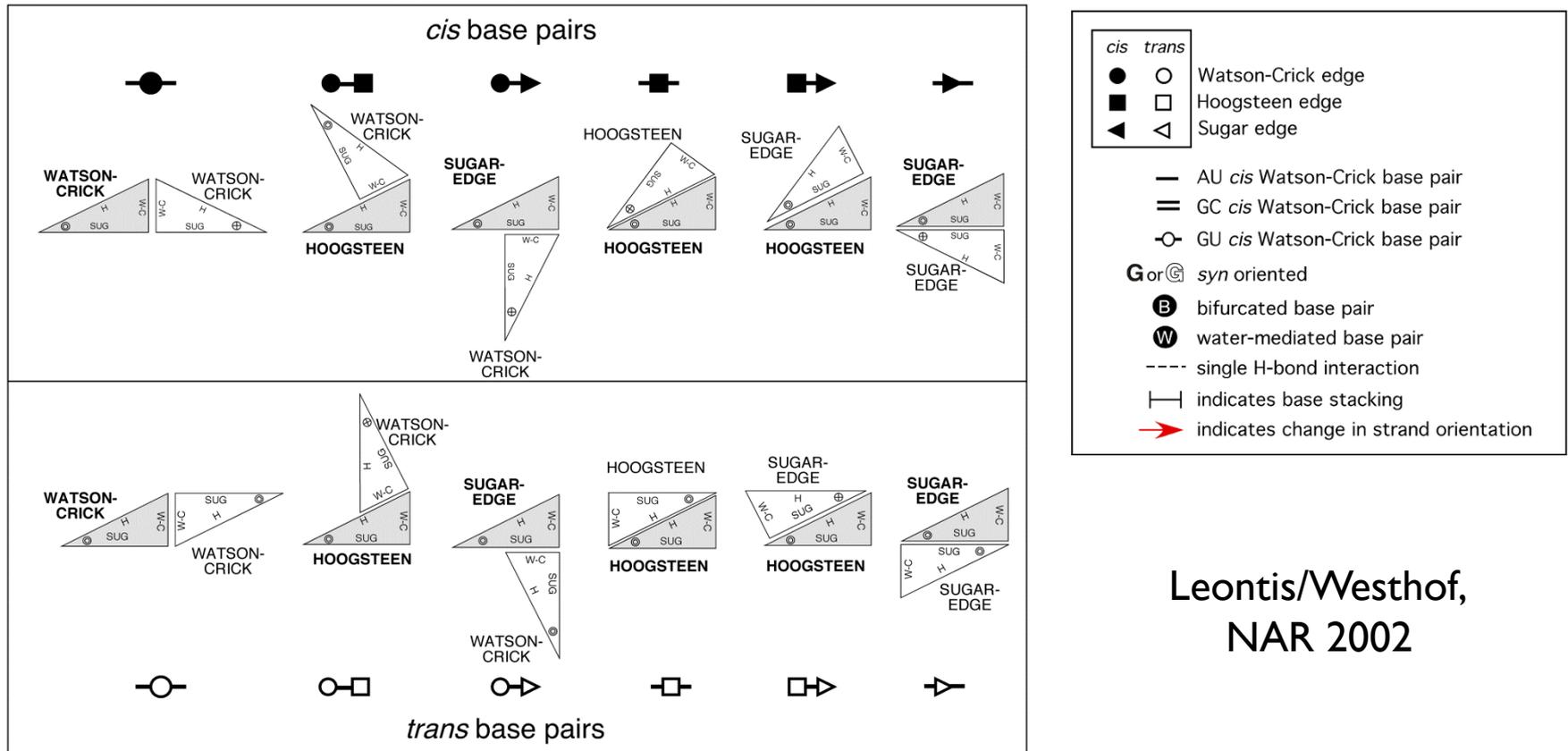
RNA nucleotides bind through edge/edge interactions.

Non canonical are weaker, but cluster into modules that are **structurally constrained**, **evolutionarily conserved**, and functionally essential.

---



# Leontis / Westhof nomenclature: A *visual grammar* for tertiary motifs



Leontis/Westhof,  
NAR 2002

+ Tools to infer base-pairs from experimentally-derived 3D models

**RNAView**, MC-Annotate...

# Automated annotation of 3D RNA models

---

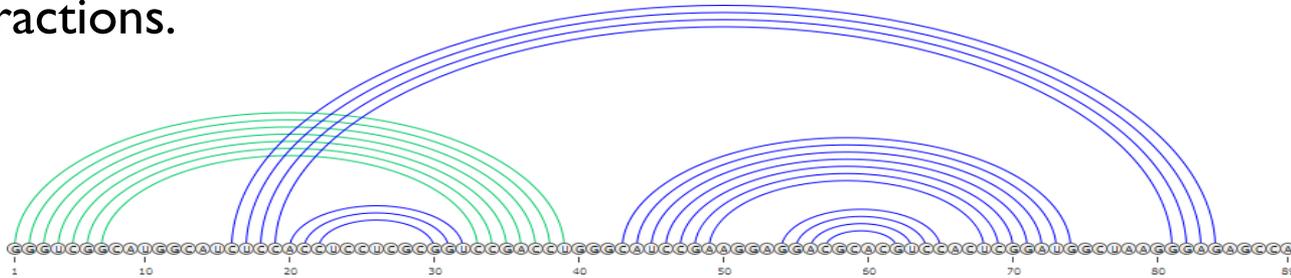
- ▶ Get from the NDB and compile (see Readme) the RNAView software\*  
<http://ndbserver.rutgers.edu/services/download/>
- ▶ Retrieve the 3IGI model from the RSCB PDB as a PDB file.
- ▶ Annotate it using RNAView (-p option) to create a RNAML file
- ▶ Visualize the output RNAML file within VARNA
- ▶ Run RNAFold (default options) on the sequence and compare the prediction with the one inferred from the 3D model.



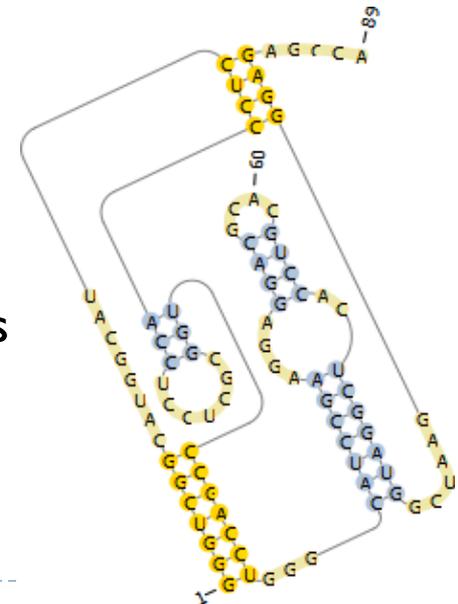
# Pseudoknots

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- ▶ Pseudoknots are complex topological models indicated by crossing interactions.



- ▶ Pseudoknots are largely ignored by computational prediction tools:
  - ▶ Lack of accepted energy model
  - ▶ Algorithmically challenging
- ▶ Yet heuristics can be sometimes efficient.
- ▶ Visualizing of secondary structure with pseudoknots is supported by:
  - ▶ PseudoViewer
  - ▶ VARNA



# Predicting and visualizing Pseudoknots

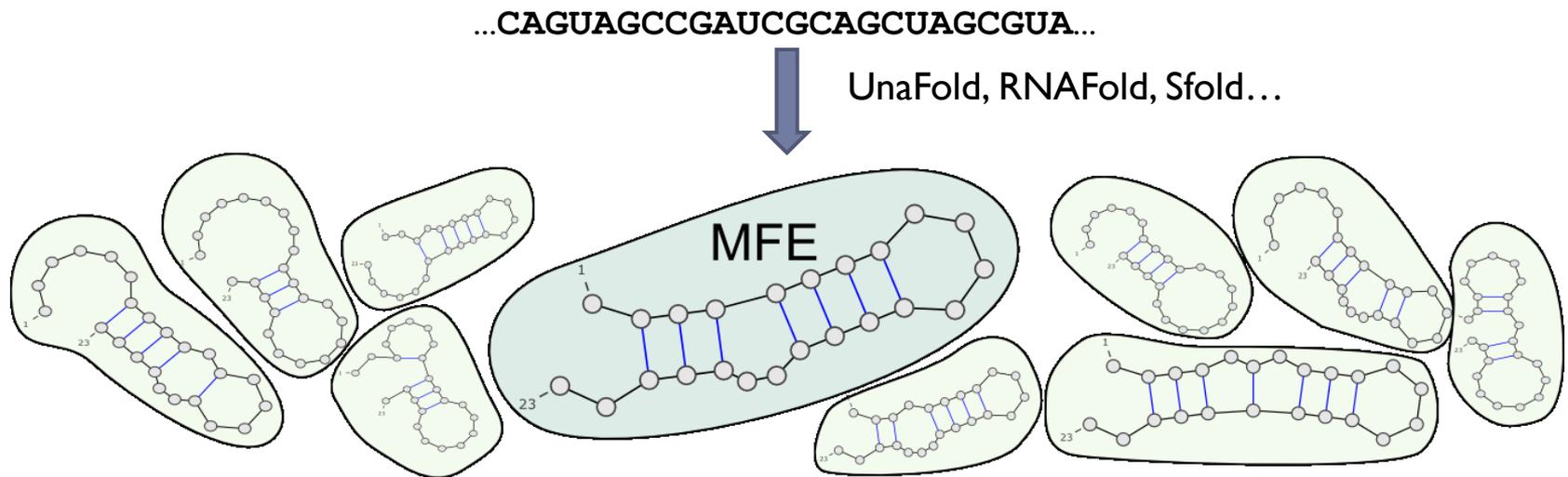
---

- ▶ Get seq./struct. data for a pseudoknot tmRNA the PseudoBase (ID: PKB210)  
<http://pseudobaseplusplus.utep.edu/>
- ▶ Visualize the structure using VARNA and the Pseudoviewer:  
<http://pseudoviewer.inha.ac.kr/>
- ▶ Fold this sequence using RNAFold and compare the result to the native structure
- ▶ Fold this sequence using Pknots-RG (*Program type: Enforcing PK*):  
<http://bibiserv.techfak.uni-bielefeld.de/pknotsrg/>



# Ensemble approaches in RNA folding

- ▶ RNA *in silico* paradigm shift:
  - ▶ From single structure, minimal free-energy folding...
  - ▶ ... to ensemble approaches.



**Ensemble diversity?** Structure likelihood? Evolutionary robustness?

Example:





# Comparative data

# RNA Alignment curation

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- ▶ Different tools for different tasks
  - ▶ ‘top down’ Structure guided modelling
    - ▶ S2S/Assemble
    - ▶ Interactive 3D modelling – edit structure based on fold predictions and manual manipulation
    - ▶ Alignments arise from RNA structure comparisons
  - ▶ ‘Bottom up’
    - ▶ Use evolutionary information (conservation patterns) to infer structural homology
    - ▶ Alignment methods like locaRNA or R-COFFEE maximise similarity in base pair contacts
    - ▶ Still need to curate/correlate with respect to other evidence for homology
  - ▶ Why curate when no structure is available
    - ▶ INFERNAL – tool to search genomes for matches to RFAM alignments
      - Functional modules, etc.



# A selection of tools ..

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- ▶ **RALEE (based on Emacs)**
  - ▶ Favourite for hardcore RNA modellers – (, ), space and delete to edit
- ▶ **4SALE**
  - ▶ Visual editor also accesses RNA alignment and folding services
- ▶ **BoulderAle: <http://boulderale.sourceforge.net/>**
  - ▶ Web based RNA alignment annotator/editor (up to 1000 nucleotides)
  - ▶ Uses VARNA for 2D visualization & KineMAGE for 3D structure
    - ▶ Stockholm file + Vienna files + GFF
  - ▶ Model 2D structure based on isostericity
  - ▶ Curate alignments to align bases that can form similar base-base interactions
- ▶ **Jalview – new kid on the block...**



# 4SALE

Cnidaria (alignment with Structures).fasta

ignore Gaps

Consensus

8101933 Alcyon C G - - U G C G C G U U C G A U U G A A

8101934 Alcyon C G - - U G C G C G U

8101948 Alcyon C G C G U G C G U G U

8101947 Alcyon C G C G U G C G U G U

8101946 Alcyon C A - - A G - - - U

8101949 Alcyon C G C G U G C G U G U

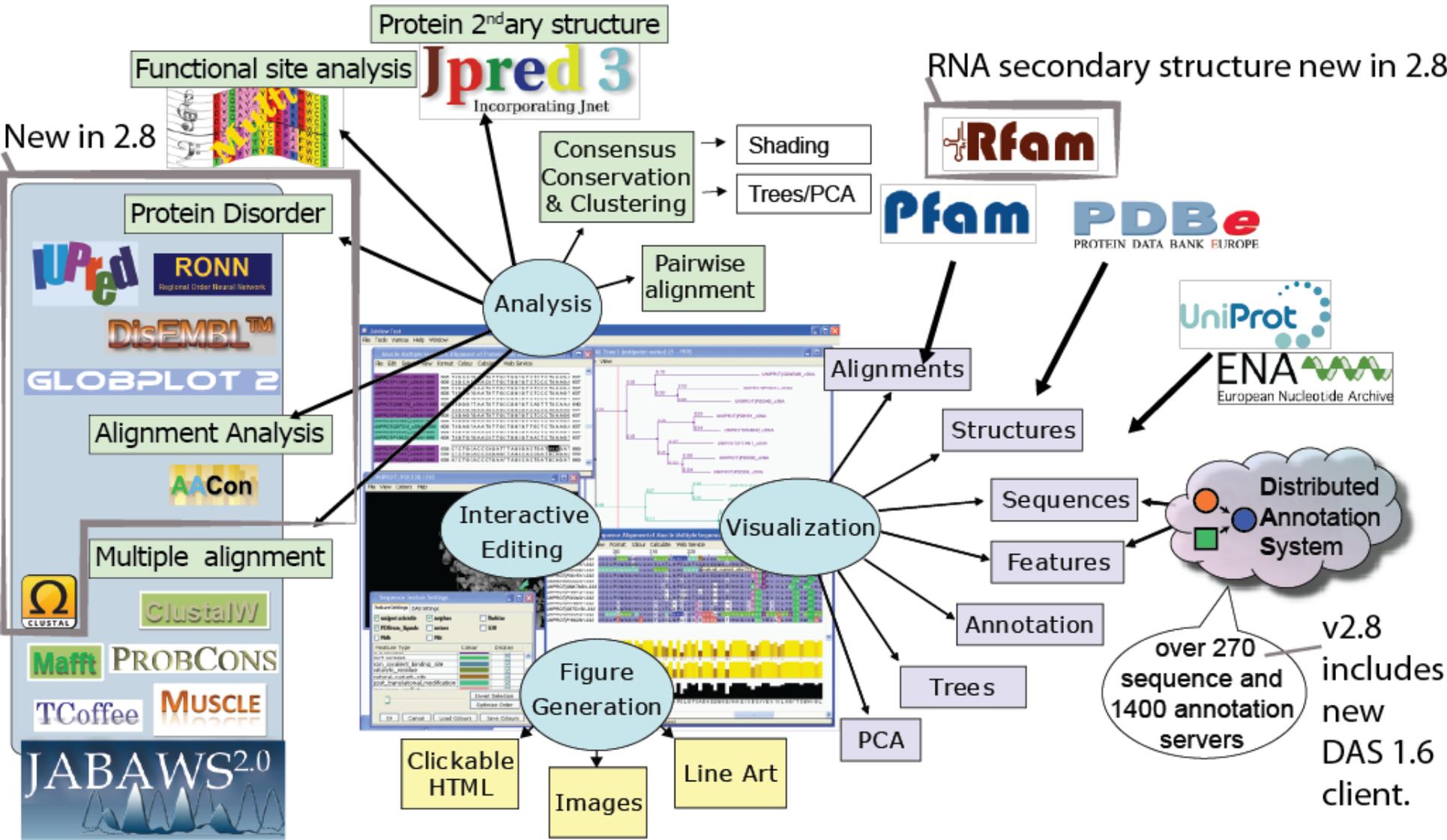
Conservation 0.0 1.0

Save Olsen Layout include Gaps

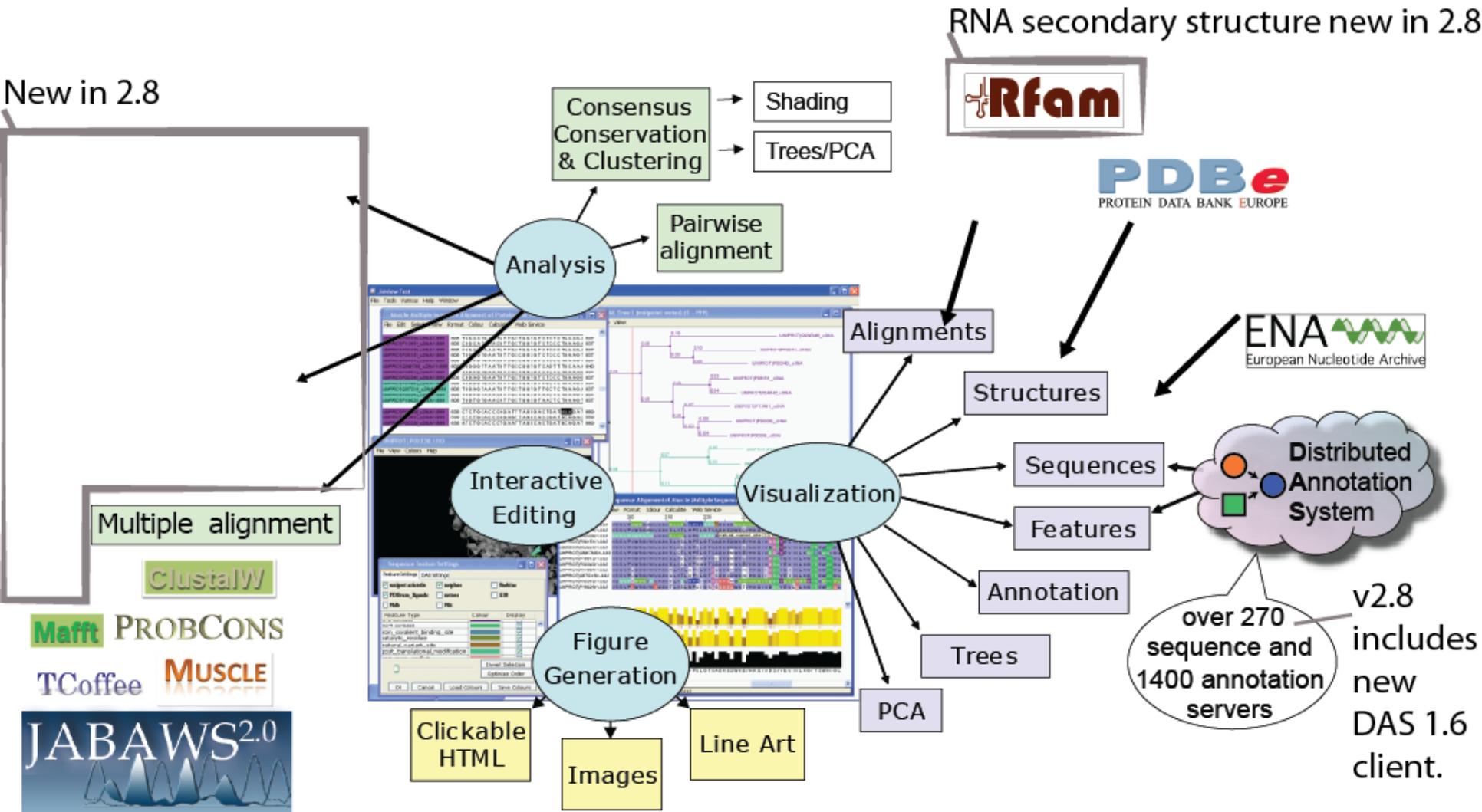
Not only selection tools, but also sequence and structure edit functions are synchronized.

ences	1	2	3	4	
01933 Alcyo		0	1	2	3
01934 Alcyo	0		1	2	3
01948 Alcyo	1	1		0	2

# Upcoming Jalview features

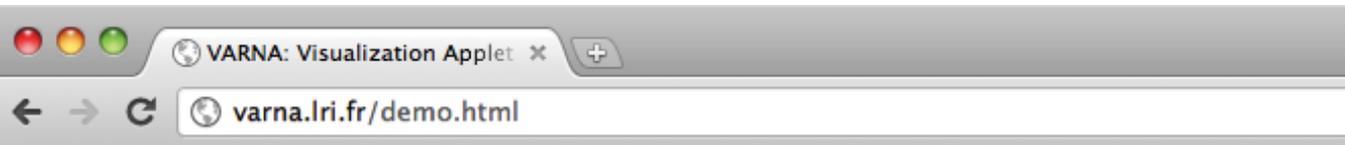
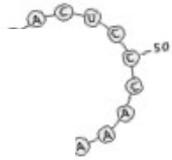


# Jalview's features relevant to RNA





# Jan Engelhardt (Uni. Leipzig)



## **VARNA: Visualization Applet for RNA**

A Java lightweight component and applet for drawing the RNA secondary structure



# RNA alignment tutorial with Locarna and Jalview

---

1. Start Development version of Jalview  
[http://www.compbio.dundee.ac.uk/users/ws-dev/development/webstart/jalview\\_1G.jnlp](http://www.compbio.dundee.ac.uk/users/ws-dev/development/webstart/jalview_1G.jnlp)
2. Import RF00162 from RFAM seed alignment
3. Select first 6 sequences in alignment, copy and paste to new alignment (shift + cmd/CTRL+V)
4. Select 'Edit->remove all gaps'
5. Add PDB sequence 2gis
6. Open locarna server page at <http://rna.informatik.uni-freiburg.de:8080/LocARNA.jsp>
7. Select/copy all 7 (ctrl+a + ctrl+c) and paste into locarna input
8. Wait a few minutes...



# Viewing the locarna results in Jalview

- ▶ Jalview doesn't support direct retrieval of LocaRNA results just yet

1. Download '[alignment]' link
2. Open in a text editor
3. Replace the lower RNA secondary structure line with the 'alifold' prediction given in the locarna output
4. Save and load into Jalview

download [\[alignment\]](#) [\[matrix\]](#) [\[tree\]](#)

...

```
CLUSTAL W --- LocARNA 1.5.2 - Local Alignment

Bacillus_liche_1      ((((((((((.....(((((((.....
Bacillus_licheni     UUUCUUAUCAAGAGCAGGCAGAGGGGA
L_innocua_6_1_11     AUUCUUAUCCAGAGU-GGUGGAGGGGA
L_innocua_7_1_10     CUCUUAUCCAGAGC-GGUAGAGGGGA
PDB_2gis_2GIS_A_     GGCUUAUCAAGAG-AGGUGGAGGGGA
Pelobacter_propi     UGCUUAUCAAGAGU-GGUGGAGGGGA
Bacillus_liche_2     CUCUUAUCCCGAGCUGGUGGAGGGGA
                    ((((((((((.....(((((((.....

Bacillus_liche_1     )))))))...)))))...
Bacillus_licheni     ---CGCUGAGAGAUAGGA-
L_innocua_6_1_11     UUUGUUCUGAAAGAUAGAGAA-
L_innocua_7_1_10     UCUCUG-AACGAUGAGAG-
PDB_2gis_2GIS_A_     ---CGUUGAAAGAUAGGCCA
Pelobacter_propi     ---GAUGAGAGGGUGUGCU-
Bacillus_liche_2     AUACCUUGAGCGAUAGAG-
                    .)))))...)))))...
                    (-66
```

```
alifold                )))))))...)))))
                    ((((((((((.....(((((((.....
```

```
.)))))...)))))...)))))... (-66
```

# LocaRNA and RNAliFold in Jalview

File Edit Select View Format Colour Calculate Web Service

Original

```
Bacillus_liche_1/1-110      UCCUUAUCAAGAGU- GGUGGAGGGACUGGCCUGUGAAACC- CGGCAA
Bacillus_licheni/1-100     UUCUUAUCAAGAGCAGGCAGAGGGACAAGCCCUGAUGAAGCC- CGGCAA
L_innocua_6_1_11/1-119     AUCUUAUCCAGAGU- GGUGGAGGGAAAUGCCCUGUGAAACC- CAGCAA
L_innocua_7_1_10/1-109     CUCUUAUCCAGAGC-
PDB_2gis_2GIS_A_/1-94     GGCUUAUCAAGAG-
Pelobacter_propi/1-107     CUCUUA
Bacillus_liche_2/1-138     CUCUUA
```

1. Right-click here and select 'Add PDB ID' under structure menu.
2. Enter '2GIS'.
3. Right click again and select 'View 2GIS' under 'View structure' menu to show structure.

RNAliFold

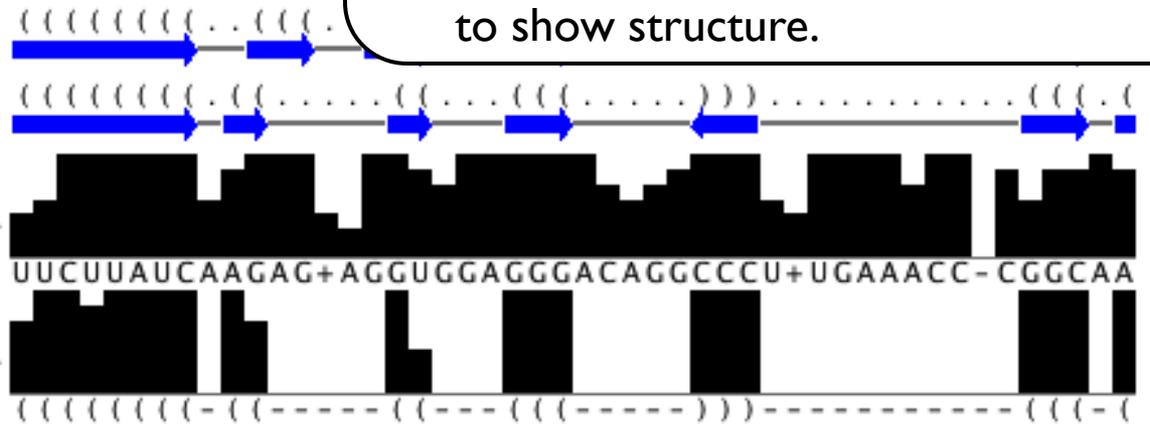
Consensus Secondary Structure

Secondary Structure

locaRNA

Consensus

StrucConsensus



Fraction of aligned WC pairs. Right-click to show pair-logo

# VARNA in Jalview

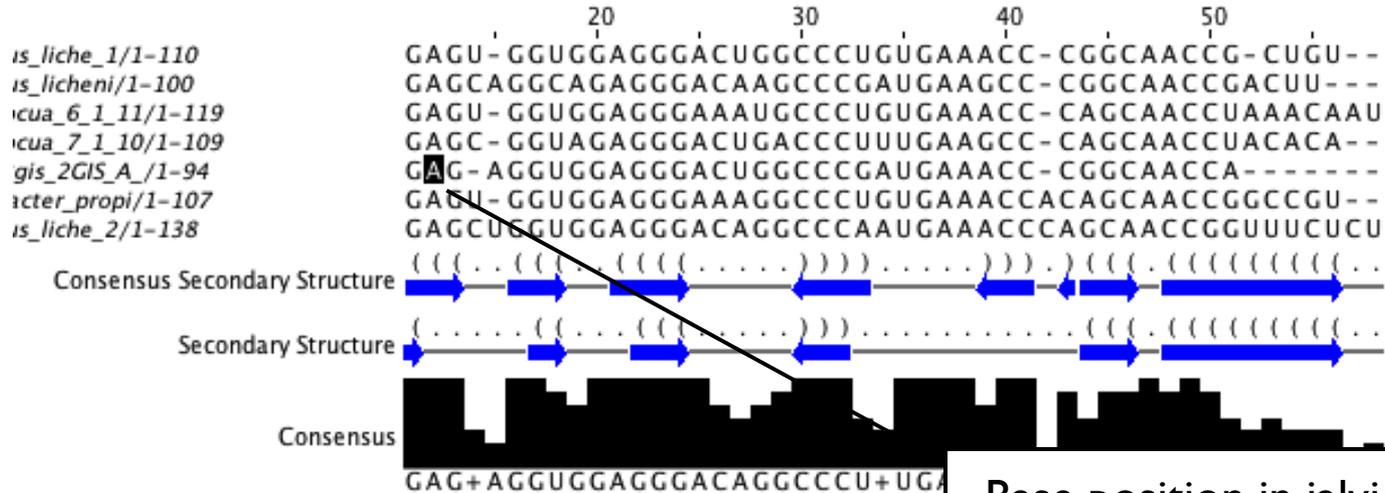
The screenshot displays the Jalview interface with the following elements:

- Sequence Alignment:** A sequence alignment of RNA sequences. The top sequence is `CCUUAUCAAGAG-AGGUGGAGGGACUGGCCCGAUGAAACC-CGGCAA`. The second sequence is `UUAUCAAGAGU-GGUGGAGGGAAAGGCCCUUGUGAAACCACAGCAA`. The third sequence is `JGAAACCCAGCAAC`.
- Context Menu:** A menu is open over the alignment with options: **Structure** (highlighted), **Hide Sequences**, **Associate Structure with Sequence**, and **View Structure**.
- Secondary Structure Visualization:** Below the alignment, four tracks are shown:
  - Consensus Secondary Structure:** A line with blue arrows indicating base pairing.
  - Secondary Structure:** A line with blue arrows indicating base pairing.
  - Consensus:** A bar chart showing the frequency of each nucleotide.
  - StrucConsensus:** A bar chart showing the frequency of each nucleotide in the structure.
- Sequence:** The consensus sequence is `UUCUUAUCAAGAG+AGGUGGAGGGACAGGCCCU+UGAAACC-CGGCAA`.
- Structure:** A line with dashed lines indicating base pairing.

On the right side, a partial view of another window shows the title `PDB_2gis_2` and buttons for `new`, `Colours`, and `Ho`. Below these buttons is a 3D ribbon model of an RNA structure.

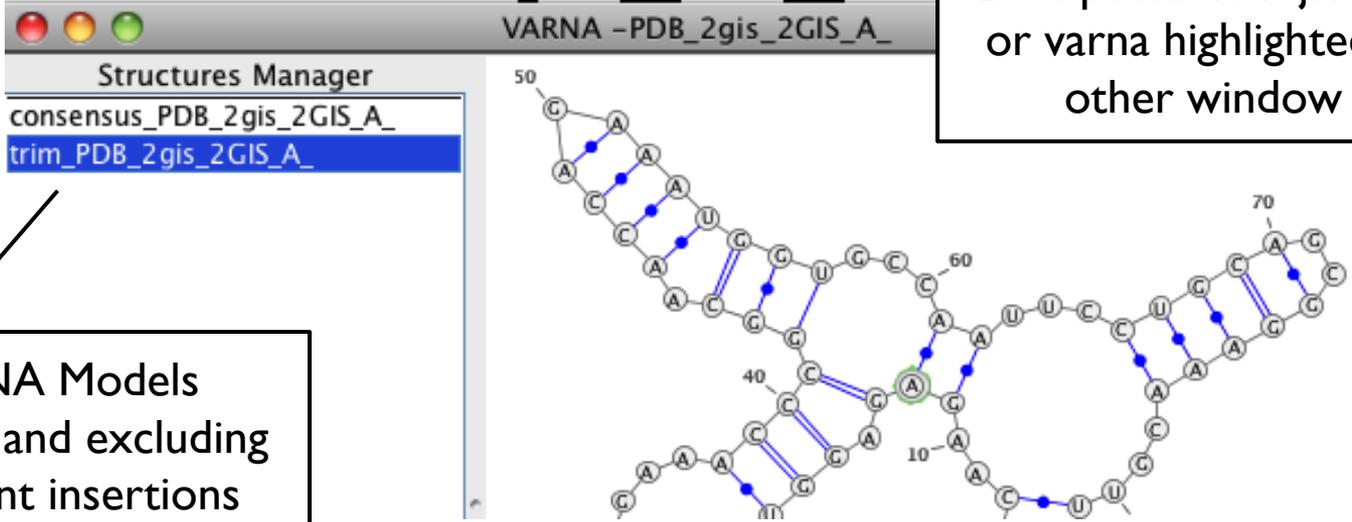


# Linked Highlighting & Selections

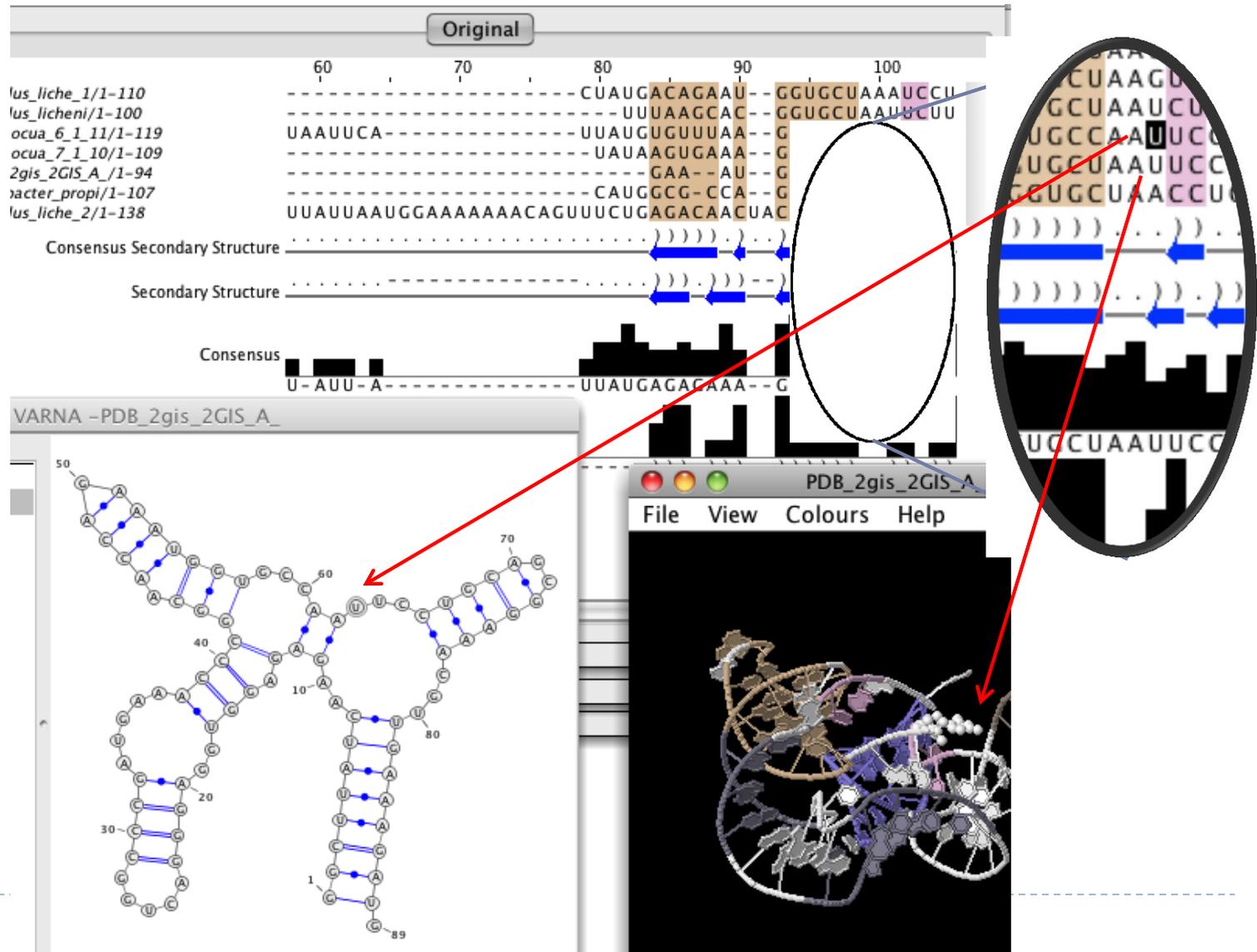


Base position in jalview  
or varna highlighted in  
other window

VARNA Models  
including and excluding  
alignment insertions



# Inspection and curation of prediction



# Summary / Discussion