

Detection and Rational Design of RNA Switches

Sebastian Will

Vienna RNA Meeting 2018

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MAY CONTAIN
TRACES OF
BIOINFORMATICS





My special perspective (on RNA research):

- **What can we learn** by applying *existing* computational tools?
e.g. exploiting the Vienna RNA package
- **Which questions could be answered** by *novel* methods?
e.g. comparing RNAs in my tool LocARNA



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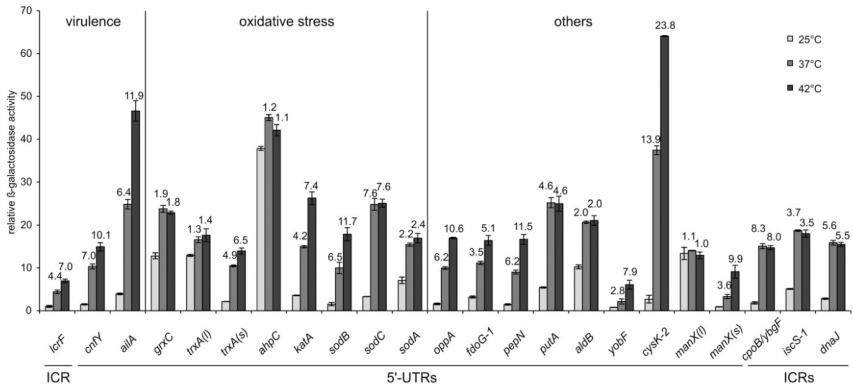


Today: What can Bioinformatics do for RNA switches?

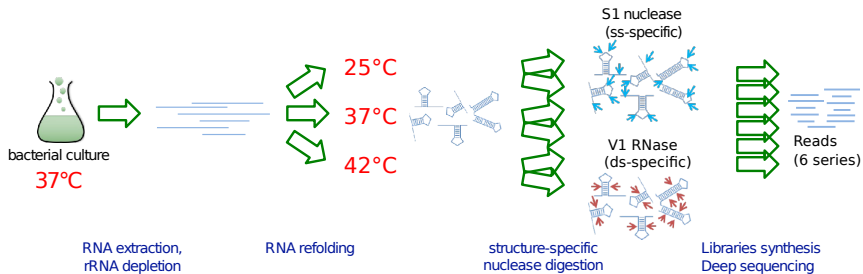
Finding Thermoswitches in *Y. Pseudotuberculosis*

with F. Righetti *et al.* **The Temperature-responsive In-vitro RNA Structure of *Yersinia Pseudotuberculosis*. *PNAS*, 2016.**

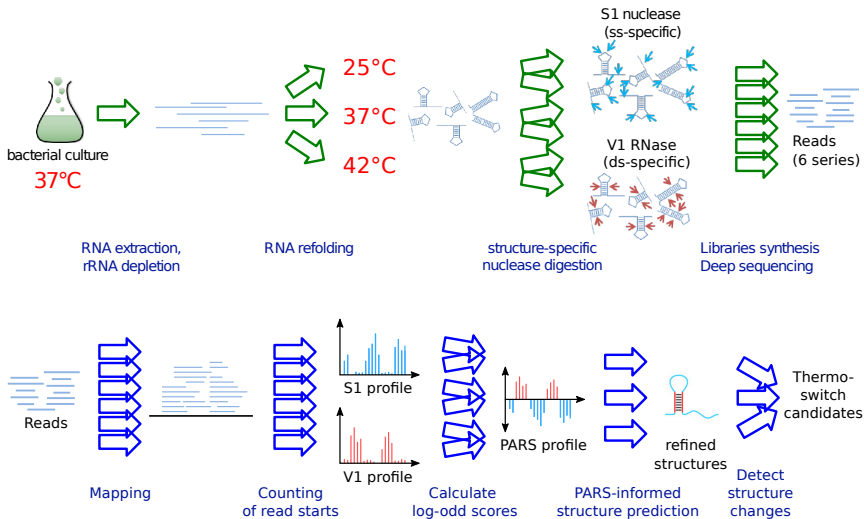
- we screened more than 1,750 RNAs at 25/37/42°C and selected *candidates* for thermoresponsive folding
- validated thermo-regulatory potential of 16 candidates



Genome-wide structure probing (PARS) at different temperatures



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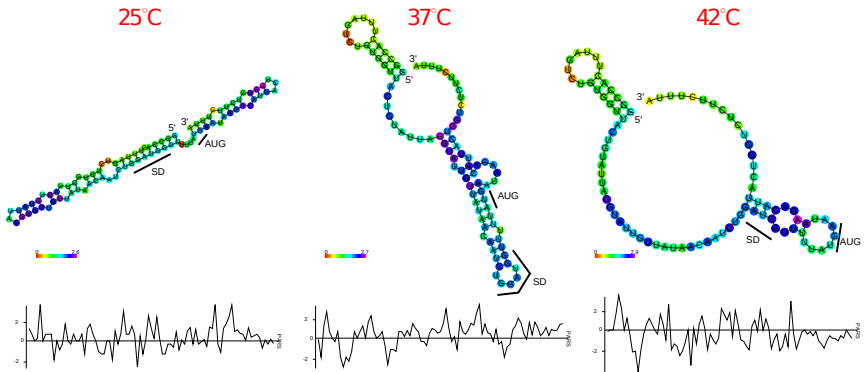
[adapted/extended from F. Righetti]

PARS at SD-region of selected RNA Thermometer candidates

Gene name	SD PARS*			PARS difference [†]		Thermal control [‡]
	25 °C	37 °C	42 °C	37–25 °C	42–25 °C	
<i>cnfY</i>	0.18	-0.37	-0.12	-0.55	-0.30	Y
<i>ailA</i>	0.09	0.61	-0.25	0.52	-0.34	Y
<i>grxC</i>	0.39	-0.56	-0.50	-0.95	-0.89	N
<i>trxA</i>	0.32	-0.53	-0.42	-0.85	-0.74	s:Y, l:N
<i>ahpC</i>	-0.60	-1.23	0.44	-0.63	1.04	N
<i>katA</i>	0.08	-0.37	-0.02	-0.45	-0.10	Y
<i>sodB</i>	0.00	-0.22	-0.29	-0.22	-0.29	Y
<i>sodC</i>	0.14	-0.08	-0.47	-0.22	-0.61	Y
<i>sodA</i>	0.09	0.07	0.02	-0.02	-0.07	N
<i>oppA</i>	0.64	-0.77	-0.13	-1.41	-0.77	Y
<i>fdoG-1</i>	0.55	-0.55	0.12	-1.10	-0.43	Y
<i>pepN</i>	0.36	-0.26	0.05	-0.62	-0.31	Y
<i>putA</i>	0.48	0.11	-0.05	-0.37	-0.53	Y
<i>aldB</i>	0.00	-0.27	0.22	-0.27	0.22	N
<i>yobF</i>	-0.08	0.01	0.02	0.09	0.10	Y
<i>cysK-2</i>	-0.04	0.07	-0.05	0.11	-0.01	Y
<i>manX</i>	-0.29	-0.11	-0.12	0.18	0.17	s:Y, l:N
<i>cpoBlybgF</i>	0.86	-0.33	-0.20	-1.19	-1.06	Y
<i>iscS-1</i>	0.43	0.16	-0.22	-0.27	-0.65	Y
<i>dnaJ</i>	0.18	-0.05	-0.10	-0.23	-0.28	Y

Results for RNA thermometer candidate *ailA*

Two candidates associated with *Y. virulence*: *ailA* and *cnfY*.

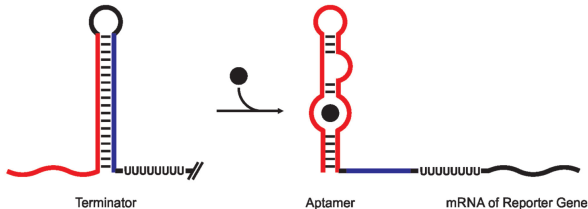


PARS profiles and PARS-guided predictions; color=entropy

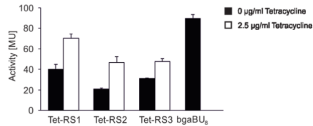
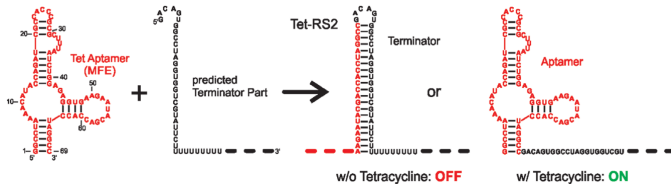
Rational Design of Riboswitches

with G. Domin *et al.* **Applicability of a computational design approach for synthetic riboswitches.** *Nucleic acids research*, 2017.

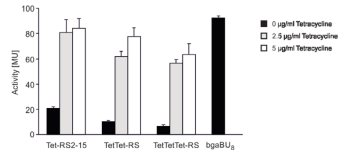
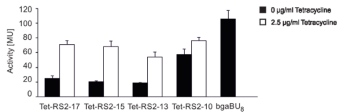
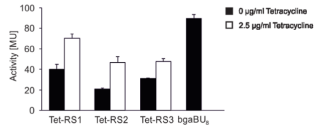
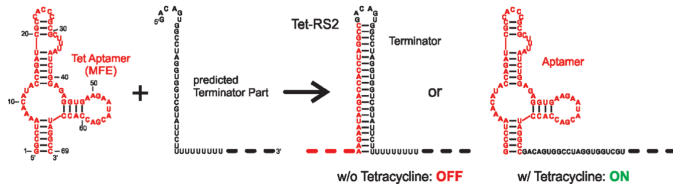
Goal: Design Riboswitch-systems to reprogram cells (here *E. coli*) to respond to small molecules (Theophylline, Tetracycline, Streptomycin)



Tetracycline-RS Designs (Activity Tests)

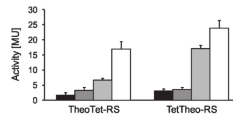
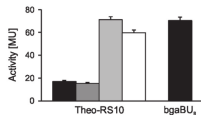
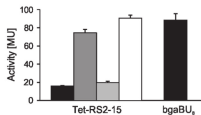
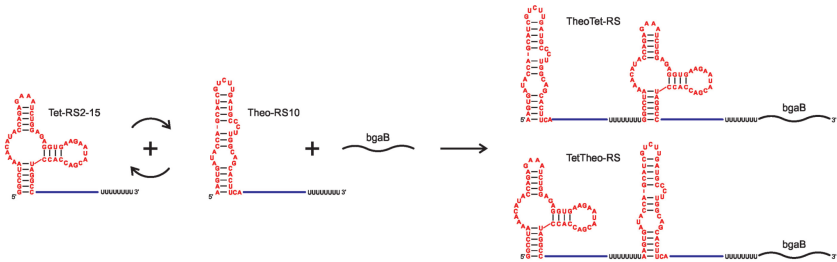


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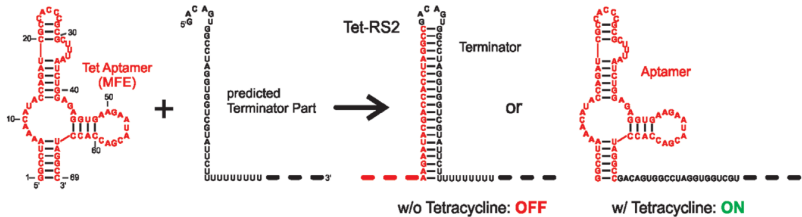
Tandem-RS Designs for AND switch

Goal: Switch ON if Theophylline AND Tetracycline are both present



Novel Method for multi-target design of RNAs

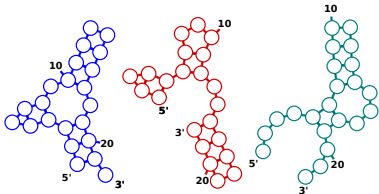
with Stefan Hammer, Wei Wang and Yann Ponty. **Fixed-Parameter Tractable Sampling for RNA Design with Multiple Target Structures.** *RECOMB*, 2018.



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Multiple structures (*targets*)

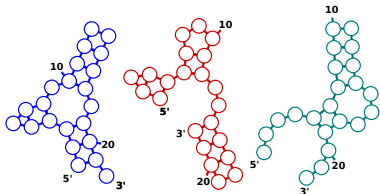


((((((.)) . ((((.))))) .)) .
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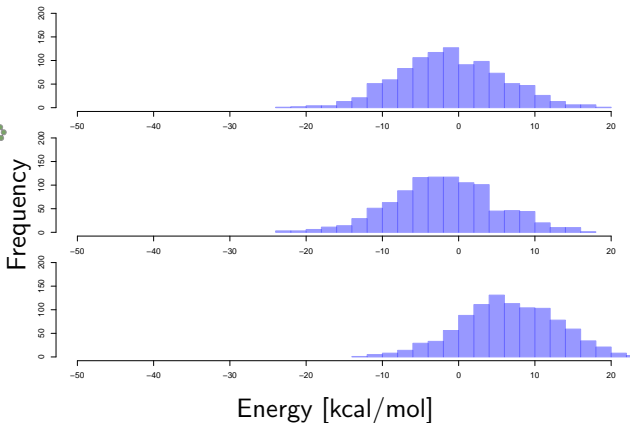
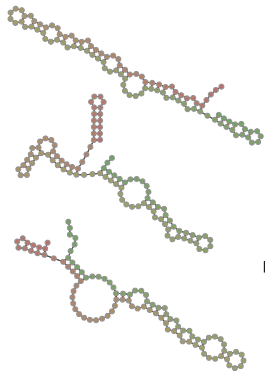
((((((.) . ((((.))) .))) .
((.) ((. . .)) . . (((. . .)))
... . ((((((. . .)))))

Task: generate seq's with *specific* properties

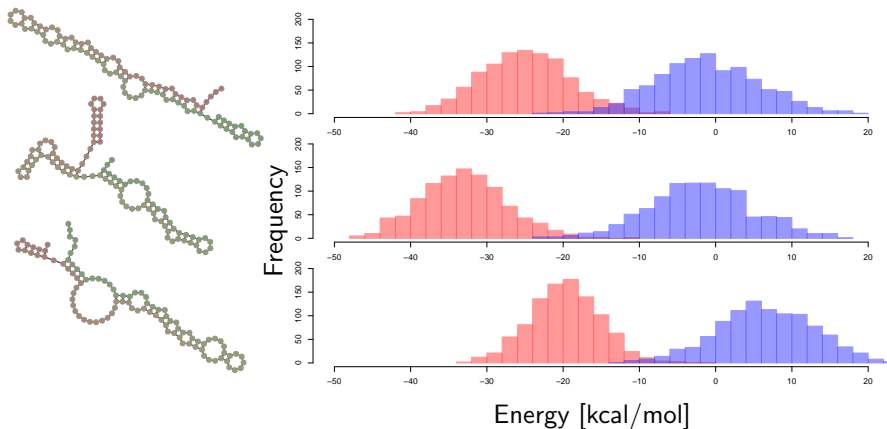
- low/specific energy for multiple structures
- specific GC content
- specific energy differences
- specific sequence/structure motifs

Approach:
defined “Boltzmann”
sampling
of RNA sequences

Multi-target design to three RNA structures

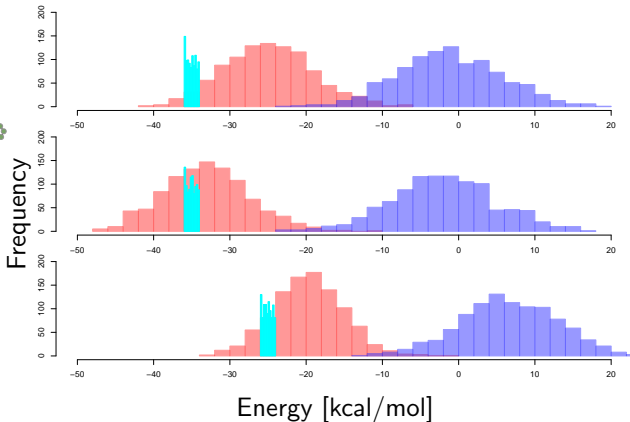
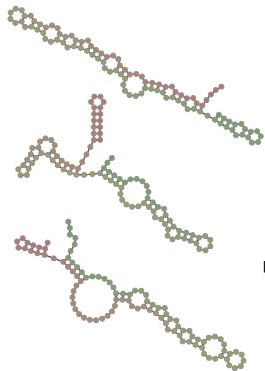


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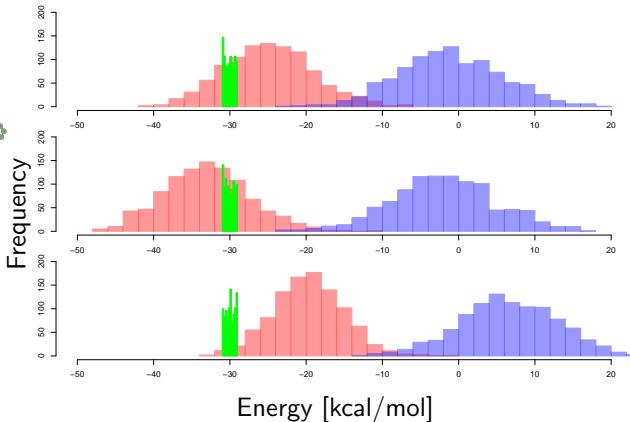
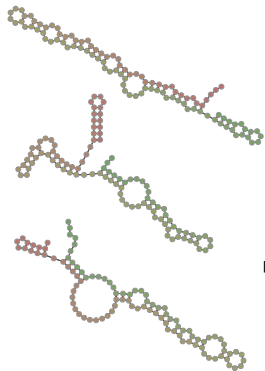
Boltzmann sample: 1000 low energy sequences; generated in seconds

Multi-target design to three RNA structures



Boltzmann sample: 1000 low energy sequences; generated in seconds
Targeted samples: 1000 highly specific sequences; in minutes

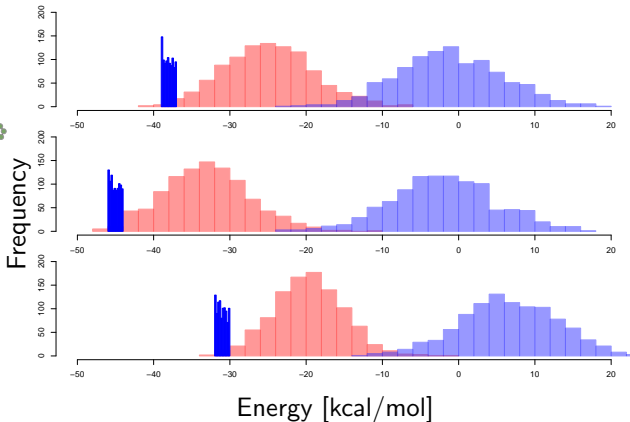
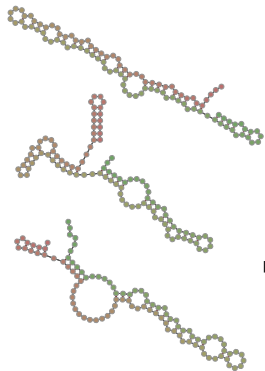
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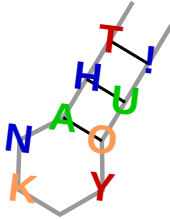
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<https://github.com/s-will/Infrared>

- **Improves quality and feasibility of RNA Design**
e.g. for designing artificial riboswitches
- **Generic system to extend RNA Design ...**
by including various desirable properties in the sampling
- **...and develop novel sampling-based tools**
 - design RNA alignments (target energies and evo-distances)
 - e.g. use to assess statistical significance
and support the detection of potential RNA switches



My co-authors / cooperation partners

Francesco Righetti, Aaron M. Nuss, Christian Twittenhoff, Sascha Beele, Kristina Urban, Stephan H. Bernhart, Peter F. Stadler, Petra Dersch, Franz Narberhaus; Gesine Domin, Sven Findeiß, Manja Wachsmuth, Mario Mörl; Stefan Hammer, Wei Wang, Yann Ponty

Team



of Ivo Hofacker at



universität
wien